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(54) Title: NOVEL INDENOISOQUINOLINES AS ANTINEOPLASTIC AGENTS

(57) Abstract

A number of indenoisoquinolines were prepared and evaluated for cytotoxicity in human cancer cell cultures and for activity vs. topoisomerase I. The two most cytotoxic indenoisoquinolines proved to be *cis*-6-ethyl-5, 6, 12, 13-tetrahydro-2, 3-dimethoxy-8, 9-(methylenedioxy)-5, 11-dioxo-11*H*-indeno[1,2-*c*] isoquinoline and *cis*-6-allyl-5, 6, 12, 13-tetrahydro-2, 3-dimethoxy-8, 9-(methylenedioxy)-5, 11-dioxo-(11*H*)indeno[1,2-*c*] isoquinoline. Two of the most potent topoisomerase I inhibitors were 6-(3-carboxy-1-propyl)-5, 6-dihydro-5, 1 Idioxo-11*H*-indeno[1,2-*c*] isoquinoline (26) and 6-ethyl-2, 3-dimethoxy-8, 9-(methylenedioxy)11*H*-indeno[1,2-*c*]isoquinolinium chloride (27). Two additional potent topoisomerase I inhibitors, 6-allyl-5, 6-dihydro-2, 3-dimethoxy-8, 9-(methylenedioxy)-5, 11-dioxo-11*H*-indeno[1,2-*c*] isoquinoline (13c) and 5, 6-dihydro-6-(4-hydroxybut-1-yl)-2, 3-dimethoxy-8, 9-methylenedioxy-5, 11 dioxo-(11*H*)indeno[1, 2-*c*]isoquinoline (19a), did not unwind DNA and did not affect topoisomerase II.

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NOVEL INDENOISOQUINOLINES AS ANTINEOPLASTIC AGENTS

Field of the Invention

The present invention relates to compositions and a method for treating
5 a patient having cancer. More specifically, the present invention is directed to novel
indenoisoquinoline derivatives and to their use in cancer therapy.

Government Rights

This invention was made with Government support under Grant No.
10 NO1-CM-67260 awarded by the National Institutes of Health. The government has
certain rights in the invention.

Background and Summary of the Invention

The control and cure of cancer represents one of our most challenging
15 health problems. The treatment of cancer can be approached by several modes of
therapy including surgery, radiation, chemotherapy or a combination of any of these
treatments. Chemotherapy continues to be an indispensable therapy for inoperable or
metastatic forms of the disease. Thus, the discovery of compounds specifically
targeting cancer cells, or the cellular mechanisms involved in the proliferation of cancer
20 cells, can provide significant advancement in the eradication or control of cancer.

The selection of compounds having effective anticancer activity is
complicated by the still limited knowledge of cancer cell biology and biochemistry.
Therefore, development of new effective anti-cancer agents remains heavily dependent
on screening of new compounds for cytotoxic activity. Antineoplastic drug candidates
25 exhibit enhanced cytotoxicity against cancer cells relative to normal cells. Methods of
screening for anticancer activity have focused on several targets: (1) the ability of a
compound to inhibit tumor growth and/or progression in animal studies; (2) inhibition
of cell growth/proliferation in cell lines of cancerous origin; and (3) inhibition of
intracellular processes necessary for the growth or propagation of cancer cells.

30 The mouse L1210 leukemia cell line was initially the preferred model
system used for screening compounds for anti-cancer activity. However, the P388
murine leukemia system was found to be more sensitive and predictive than L1210

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The mouse L1210 leukemia cell line was initially the preferred model system used for screening compounds for anti-cancer activity. However, the P388 murine leukemia system was found to be more sensitive and predictive than L1210 leukemia system; it has been used as a primary screen during the past decade.

5 Systematic screening for compounds exhibiting toxicity to these two cell lines has resulted in the isolation of a large number of active natural products. However, the anticancer activities of these compounds were predominantly for leukemia, lymphoma and a few rare tumors. Low clinical efficacy, or the lack of clinical efficacy of known chemotherapeutics against slower growing solid tumors, is a serious concern.

10 Considering the diversity of cancer in terms of cell type, morphology, growth rate and other cellular characteristics, the U.S. National Cancer Institute (NCI) has developed a "disease-oriented" approach to anticancer activity screening (M.R. Boyd, in "Principle of Practice of Oncology" J.T. Devita, S. Hellman, S.A. Rosenberg (Eds.) Vol. 3, PPO Update, No. 10, 1989). This *in vitro* prescreening system is based
15 on the measurement of anticancer cytotoxicity against human cancer cell line panels consisting of approximately 60 cell lines of major human cancers (including leukemia and slower growing tumor cells such as lung, colon, breast, skin, kidney, etc.) and is referred hereinafter as "COMPARE" screening. An important advantage of the new
20 *in vitro* screening panels is the opportunity to facilitate identification of compounds that are selectively more cytotoxic to cells of certain types of cancers, thus increasing the ability to select compounds for further study with respect to specific diseases.

25 The compounds of the present invention were screened for antineoplastic activity using the COMPARE screening methodology. The results demonstrate that the compounds are antineoplastic agents for use in treating human cancers.

Anticancer agents are known to act through a variety of mechanisms to destroy or inhibit the proliferation of cancer cells. For example, some agents are antimetabolites which act as false substrates in the biochemical processes of cancer cells. One compound which has this mechanism of action is methotrexate, an analog
30 of folic acid, which functions in part by binding to dihydrofolate reductase, thereby preventing the formation of guanine and adenine from the folic acid precursor

molecule. Thus, methotrexate inhibits the ability of cancer cells to construct DNA by inhibiting the proper metabolism of folic acid.

Other anticancer agents act by alkylating DNA strands, thereby producing defects in the normal double helical structure of the DNA molecule. This 5 alkylation may cause the formation of breaks and inappropriate links between (or within) strands of DNA. Such disruption of the DNA structure, if not repaired by intracellular repair mechanisms, impairs the cell's ability to replicate it's DNA.

Examples of alkylating anticancer agents are cyclophosphamide and chlorambucil.

Some anticancer agents target the intracellular mechanisms involved in 10 replication of the DNA strand itself. Replication of a cell's genetic material requires a means to pull the DNA double helix apart into two strands. This separation is typically accomplished by the enzyme topoisomerase I. Disruption of the function of this enzyme results in DNA strand breaks in cells that are dividing, thereby causing the death of the dividing cell. Because cancer cells grow and reproduce at a much 15 faster rate than normal cells, they are more vulnerable to topoisomerase inhibition than are normal cells. Thus, agents that inhibit topoisomerase I are known to be potent anticancer agents. The drug camptothecin was shown to be an inhibitor of topoisomerase I and a potent anticancer agent; unfortunately, camptothecin also produced toxic side effects. The search for potent inhibitors of topoisomerase I with 20 lessened toxicity to normal cells continues.

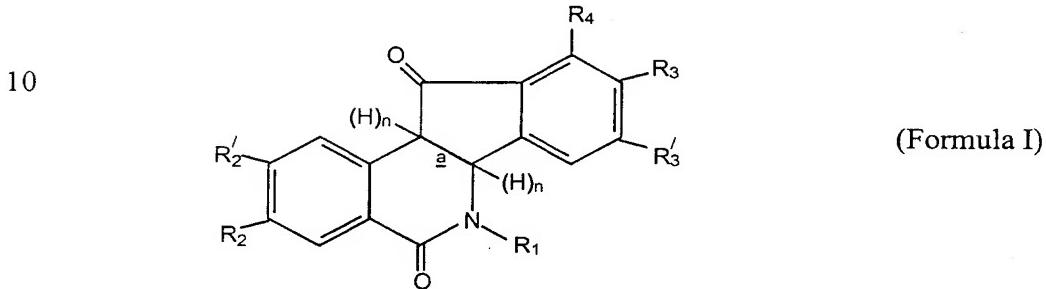
Many of the compounds of the present invention caused inhibition of topoisomerase I, to varying extents. Therefore, it appears that some of the growth inhibition demonstrated through COMPARE testing occurs through this mechanism of action. However, several of the indenoisoquinolines of the present invention were 25 surprisingly potent cell growth inhibitors even though their inhibitory effects on topoisomerase I were relatively small in comparison to other agents tested. These data demonstrate that the novel indenoisoquinolines of the present invention cause inhibition of cell growth, at least in part, through another mechanism of action besides inhibition of topoisomerase I. The present invention describes novel 30 indenoisoquinoline compounds, many of which are potent inhibitors of topoisomerase I, and are useful as anticancer agents. Further, the present invention describes novel

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indenoisoquinoline compounds which are potent inhibitors of cell growth, and are thus potent anticancer agents.

5 Detailed Description of the Invention

The compounds of this invention are represented by the general formula:



15

wherein the group designated R₁ is hydrogen, formyl, phenyl, phenyl substituted with C₁-C₆ alkoxy or C₁-C₆ alkyl, or R₁ is a group -(CH₂)_mZ, wherein m is 1-6 and Z is selected from the group consisting of hydrogen, hydroxy, carboxy, formyl, C₁-C₆ alkyl, carbo-(C₁-C₆ alkoxy), C₂-C₆ alkenyl, phenyl, C₁-C₆ alkylamino, and C₁-C₆ hydroxyalkylamino;

R₂, R_{2'} and R₄ are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₂-C₆ alkenyl, C₁-C₆ alkoxy, phenoxy and benzyloxy, or R₂ and R_{2'} taken together form a group of the formula -OCH₂O-;

R₃ and R_{3'} are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy, or R₃ and R_{3'} taken together form a group of the formula -OCH₂O-;

wherein n = 1 or 0, and bond a is a single bond when n = 1, and bond a is a double bond when n = 0;

provided that when R₂, R_{2'}, R₄, R₃ and R_{3'} are hydrogen, Z is not C₁-C₆ hydroxyalkylamino; and

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further provided that when R₁ is methyl, R₃ and R_{3'} are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy.

- In one preferred embodiment of the compounds of this invention of
 5 Formula I, the protons on the carbon atoms at fusion bond a are in a cis-configuration across bond a.

In one embodiment of the present invention, the compound of Formula I has the following substituents: R₁ is -(CH₂)_mO4 and m is 3-6; n is zero (0) and a is a double bond; and R₂, R_{2'}, R₃, R_{3'} and R₄ are hydrogen.

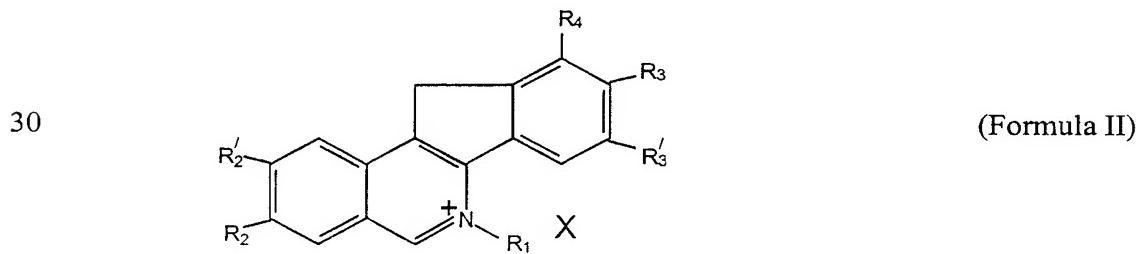
- 10 In another embodiment of the present invention, the compound of Formula I has the following substituents: R₁ is C₂-C₄ alkyl or C₂-C₄ alkenyl; R₂ and R_{2'} are C₁-C₄ alkoxy; R₃ and R_{3'} taken together form a group of the formula -OCH₂O-; and R₄ is hydrogen.

- 15 Another embodiment of the present invention includes the compound of Formula I wherein: R₁ is (CH₂)_mOH and m is 3-6; n is zero (0) and a is a double bond; R₂ and R_{2'} are C₁-C₃ alkoxy; R₃ and R_{3'} taken together form a group of the formula - OCH₂O-; and R₄ is hydrogen.

- 20 A further embodiment of the present invention includes the compound of Formula I wherein: R₁ is C₁-C₃ alkyl or C₂-C₄ alkenyl; n is one (1) and a is a single bond; R₃ and R_{3'} taken together form a group of the formula - OCH₂O-; and R₄ is hydrogen.

Another embodiment of the present invention includes the compound of Formula I wherein: R₁ is -(CH₂)_mCOOH and m is 1-4; n is zero (0) and a is a double bond; and R₂, R_{2'}, R₃, R_{3'} and R₄ are hydrogen.

- 25 Other compounds of the present invention are represented by the following formula:



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wherein R₁ is phenyl or phenyl substituted with C₁-C₆ alkoxy or C₁-C₆ alkyl, or R₁ is a group -(CH₂)_mZ wherein m is 1-6 and Z is selected from the group consisting of hydrogen, hydroxy, carboxy, formyl, C₁-C₆ alkyl, carbo-(C₁-C₆ alkoxy), C₂-C₆ alkenyl, phenyl, C₁-C₆ alkylamino, and C₁-C₆ hydroxyalkylamino, provided that
5 when Z is hydrogen, m is 2-6;

R₂, R₂' and R₄ are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₂-C₆ alkenyl, C₁-C₆ alkoxy, phenoxy and benzyloxy, or R₂ and R₂' taken together form a group of the formula -OCH₂O-;

10 R₃ and R₃' are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy, or R₃ and R₃' taken together form a group of the formula -OCH₂O-; and

wherein X is a pharmaceutically acceptable anion.

A "pharmaceutically acceptable anion" is defined as any non-toxic mono-, di-, or trivalent anions. Exemplary of such are Br⁻, Cl⁻, SO₄⁻², PO₄⁻³, acetate,
15 CO₃⁻² and HCO₃⁻. It is understood that the stoichiometry of the salts of Formula II are dependent on the valence of the anion component and the ratio of cationic to anionic components is such as to provide a neutral salt.

In one embodiment of the present invention, a compound of Formula II has the following substituent groups: R₁ is C₁-C₄ alkyl; R₂ and R₂' are C₁-C₃ alkoxy;
20 R₃ and R₃' taken together form a group of the formula -OCH₂O-; and R₄ is hydrogen.

The present invention further provides pharmaceutical formulations comprising an effective amount of an indenoisoquinoline compound of this invention for treating a patient having cancer. As used herein, an effective amount of the indenoisoquinoline compound is defined as the amount of the compound which, upon
25 administration to a patient, inhibits growth of cancer cells, kills malignant cells, reduces the volume or size of the tumors or eliminates the tumor entirely in the treated patient.

The effective amount to be administered to a patient is typically based on body surface area, patient weight, and/or patient condition. The interrelationship
30 of dosages for animals and humans (based on milligrams per meter squared of body surface) is described by Freireich, E.J., et al., *Cancer Chemother. Rep.* 1966, 50 (4), 219. Body surface area may be approximately determined from patient height and

weight (see e.g., Scientific Tables, Geigy Pharmaceuticals, Ardley, New York, pages 537-538 (1970)). An effective amount of the indenoisoquinoline compounds of the present invention is defined as any amount useful for inhibiting the growth of (or killing) cancer cells in a patient. Typically, such effective amounts range from about 5 mg/kg to about 500 mg/kg, more preferably from about 5 mg/kg to about 250 mg/kg, and most preferably about 5 to about 150 mg/kg. Effective doses will also vary, as recognized by those skilled in the art, dependent on route of administration, excipient usage and the possibility of co-usage with other therapeutic treatments including other anti-tumor agents, and radiation therapy.

10 The pharmaceutical formulation may be administered via the parenteral route, including subcutaneously, intraperitoneally, intramuscularly and intravenously. Examples of parenteral dosage forms include aqueous solutions of the active agent, in isotonic saline, 5% glucose or other well-known pharmaceutically acceptable liquid carrier. In one preferred aspect of the present embodiment, the indenoisoquinoline 15 compound is dissolved in a saline solution containing 5% dimethyl sulfoxide and 10% Cremphor EL (Sigma Chemical Company). Additional solubilizing agents such as cyclodextrins, which can form specific, more soluble complexes with the present indenoisoquinoline compounds, or other solubilizing agents well-known to those familiar with the art, can be utilized as pharmaceutical excipients for delivery of the 20 indenoisoquinoline compounds.

 The present compound can also be formulated into dosage forms for other routes of administration utilizing well-known methods. The pharmaceutical compositions can be formulated, for example, in dosage forms for oral administration in a capsule, a gel seal or a tablet. Capsules may comprise any well-known 25 pharmaceutically acceptable material such as gelatin or cellulose derivatives. Tablets may be formulated in accordance with conventional procedure by compressing mixtures of the active indenoisoquinoline and solid carriers, and lubricants well-known to those familiar with the art. Examples of solid carriers include starch, sugar and bentonite. The compounds of the present invention can also be administered in a 30 form of a hard shell tablet or capsule containing, for example, lactose or mannitol as a binder and conventional fillers and tableting agents.

The examples provided illustrate various embodiments of Applicants' invention, and are not intended to in any way limit the scope of the invention as set forth in this specification and claims.

The synthesis of an indenoisoquinoline **1** has been previously reported.

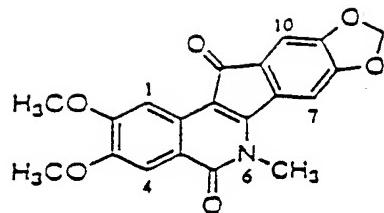
- 5 Compound **1** was subsequently found to be cytotoxic in human cancer cell cultures. More recently, a COMPARE analysis indicated that the cytotoxicity profile of **1** is similar to that of the topoisomerase I inhibitors camptothecin and saintopin. When tested for activity against topoisomerase, compound **1** was in fact found to induce DNA cleavage in the presence of topoisomerase I. However, the cleavage site
10 specificity differed from that of camptothecin in that compound **1** did not cleave at all of the sites characteristic of camptothecin, while some DNA cleavage sites were unique to compound **1**. In addition, compound **1** did not produce detectable DNA unwinding, suggesting that in contrast to other non-camptothecin topoisomerase inhibitors, it is not a DNA intercalator. The present invention describes the
15 development of new topoisomerase I inhibitors and potential anticancer agents which have been developed based upon the activities associated with compound **1**.

Chemistry

- A number of indenoisoquinolines **3-8** lacking the methylenedioxy and
20 methoxy substituents of **1** were synthesized by reacting commercially available benz[*d*]indeno[1,2-*b*]pyran-5,11-dione (**2**) with various primary amines (Scheme 1). The reactions were carried out at room temperature in chloroform and the yields were generally high.

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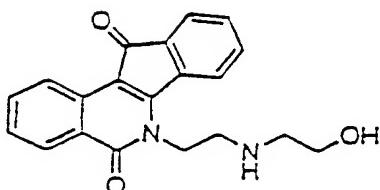
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- 3 R = CH_2CH_3
- 4 R = $CH_2CH_2CH_3$
- 5 R = cyclopropyl
- 6 R = CH_2COOCH_3
- 7 R = $CH_2CH_2CH_2CH_2OH$
- 8 R = $CH_2CH_2CH_2CH_2CH_2OH$

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Scheme 1.

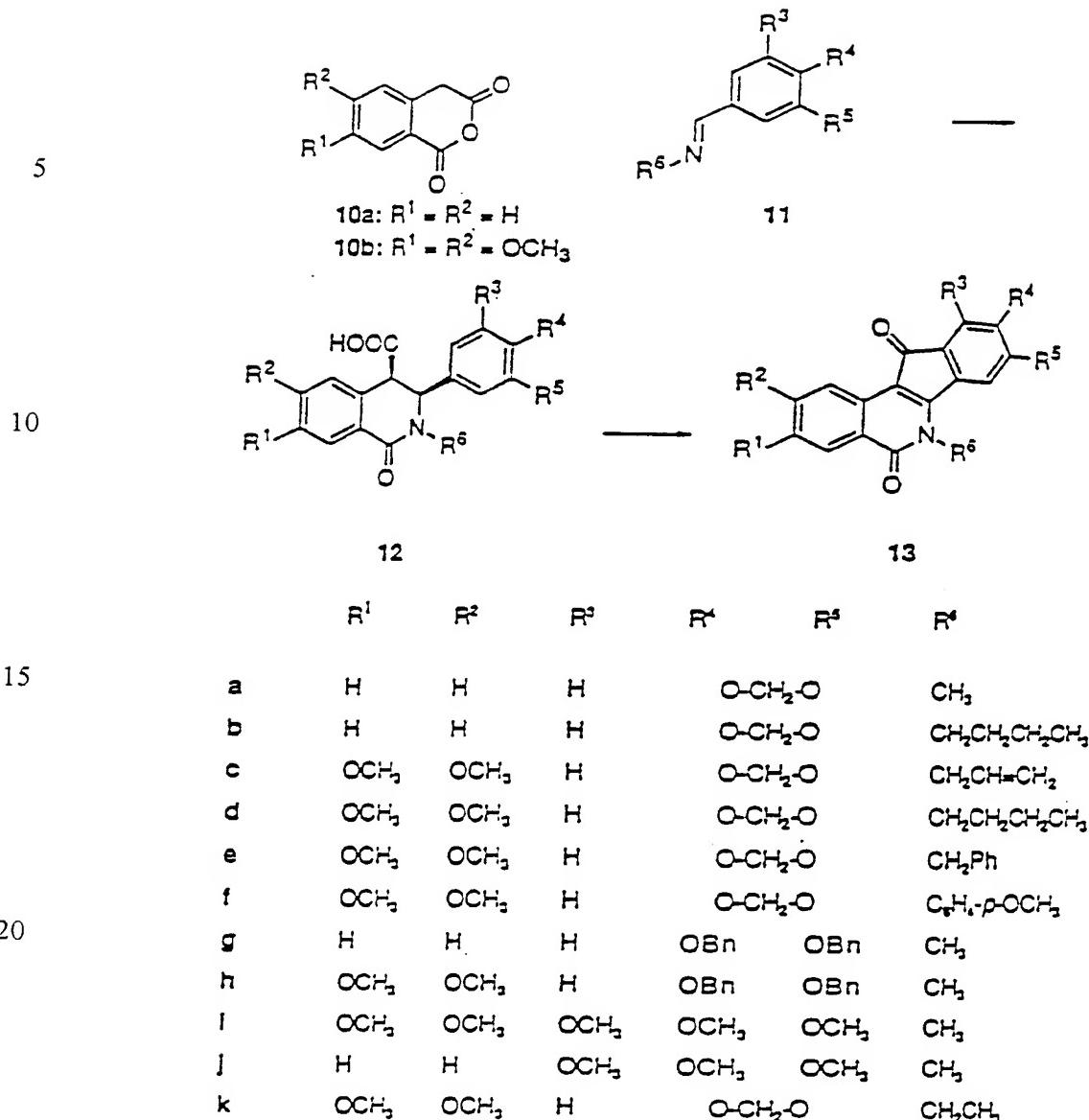
-10-

In order to accommodate additional substituents on the two aromatic rings of the indenoisoquinoline system, an alternative synthesis was executed which was based on the condensation of Schiff bases **11** with homophthalic anhydrides **10** to afford cis substituted isoquinolones **12**, followed by conversion to the desired products **13** in the presence of thionyl chloride (Scheme 2). Using this method, a series of eleven additional indenoisoquinolines **13a-13k** were synthesized. These compounds incorporate a variety of substituents at C-2, C-3, N-6, C-8, C-9, and C-10 of the ring system.

A modification of this route was carried out in order to synthesize compounds containing an alcohol group at the end of an alkyl chain located at N-6 (Scheme 3). Treatment of 4-amino-1-butanol (**14a**) or 5-amino-1-pentanol (**14b**) with *tert*-butyldimethylsilyl chloride according to the procedure of Corey and Venkateswarlu (*J. Am. Chem. Soc.* 1972, 94, 6190-6191) afforded the corresponding protected intermediates **15a** and **15b**. The imines **17a** and **17b** were synthesized by treating the *O*-TBDMS protected aminols **15a** and **15b** with piperonal (**16**) in chloroform in the presence of anhydrous magnesium sulfate. Condensation of the Schiff bases **17a** and **17b** with 4,5-dimethoxyhomophthalic anhydride (**10b**) afforded the cis 3,4-disubstituted isoquinolones **18a** and **18b**. The cis stereochemistry of **18a** and **18b** was confirmed by 6 Hz coupling constant observed for the C-3 and C-4 methine signals. Treatment of **18a** or **18b** with thionyl chloride resulted in deprotection of the terminal alcohol, allowing a Friedel-Crafts reaction to form the five-membered ring, and dehydrogenation to afford **19a** and **19b**.

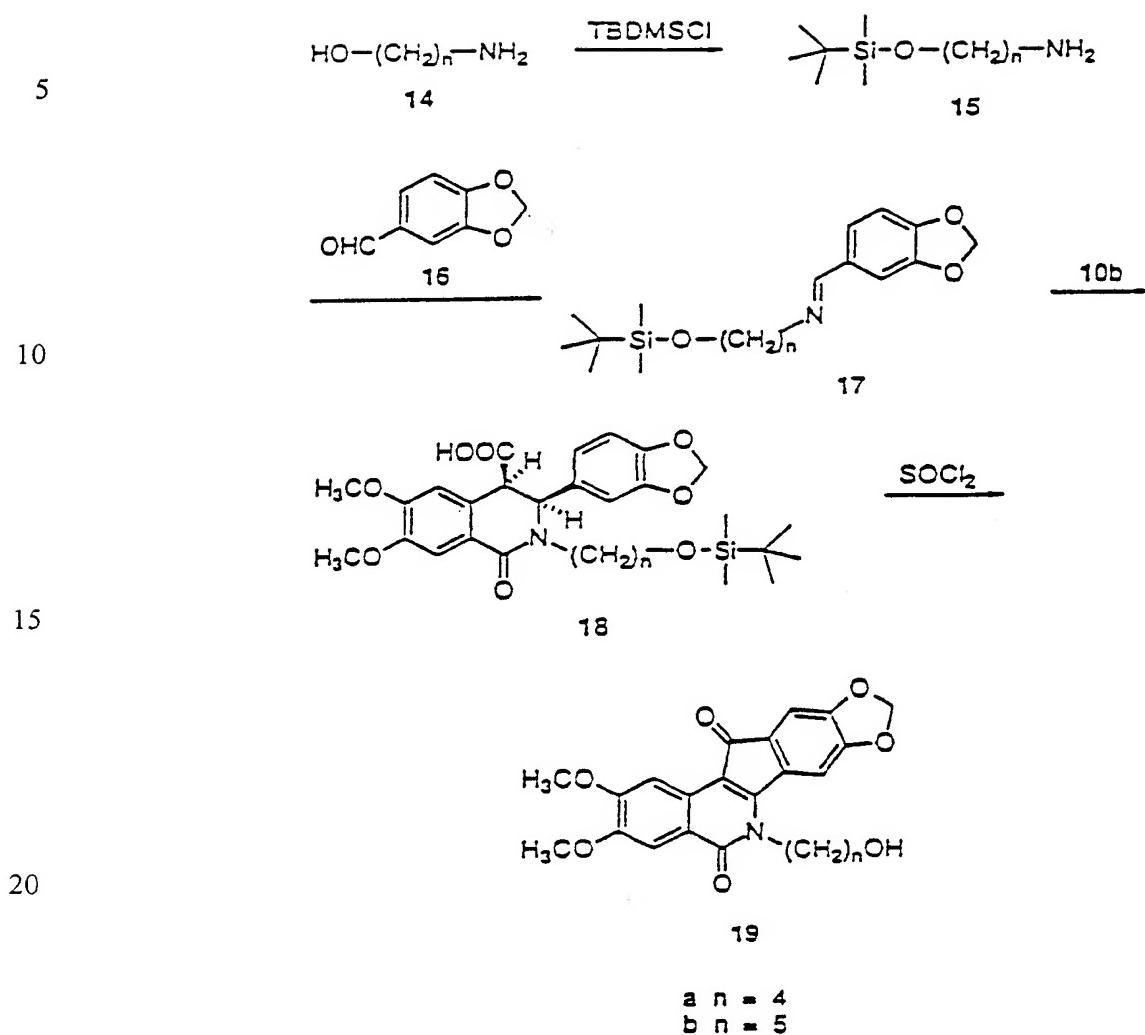
Several dihydro derivatives **20-23** were also prepared (Scheme 4). The syntheses of **20** and **23** were carried out as described previously. Compounds **21** and **22** were prepared by treatment of the acids **12k** and **12c** with Eaton's reagent (10% P₂O₅, in methanesulfonic acid). Treatment of **21** with borane-tetrahydrofuran complex in refluxing THF for 1 hour resulted in reduction of the ketone to afford **24**. When **21** was treated with the same reagent in refluxing THF for 12 hours, reduction of both the ketone and amide carbonyls occurred to yield **25**. The stereochemistry of the hydroxyl group results from the approach of the reducing reagent to the less sterically hindered, convex surface of the indenoisoquinoline **21**.

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Scheme 2.

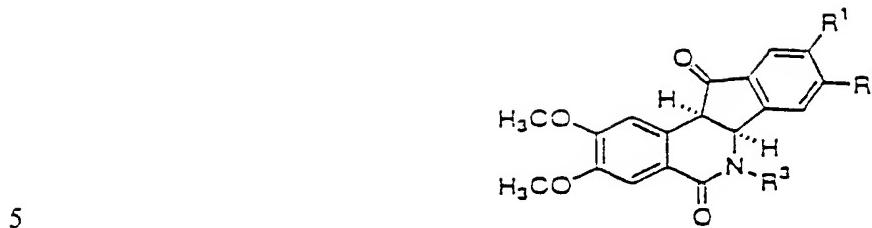
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Scheme 3.

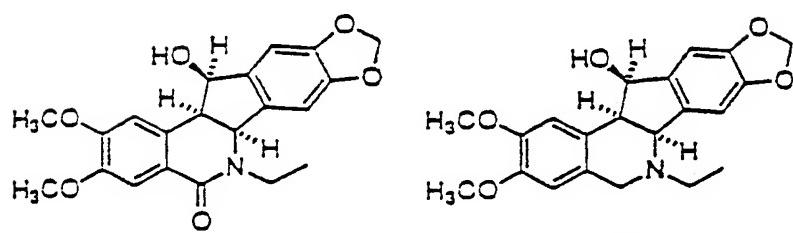
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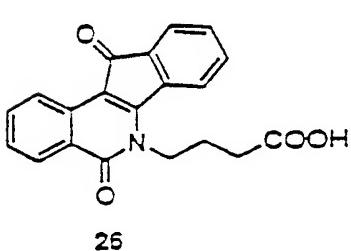


- 20 $R^1, R^2 = OCH_2O; R^3 = CH_3$
 21 $R^1, R^2 = OCH_2O; R^3 = CH_2CH_3$
 22 $R^1, R^2 = OCH_2O; R^3 = CH_2CH=CH_2$
 23 $R^1 = OSO_2CH_3; R^2 = OCH_3; R^3 = CH_3$

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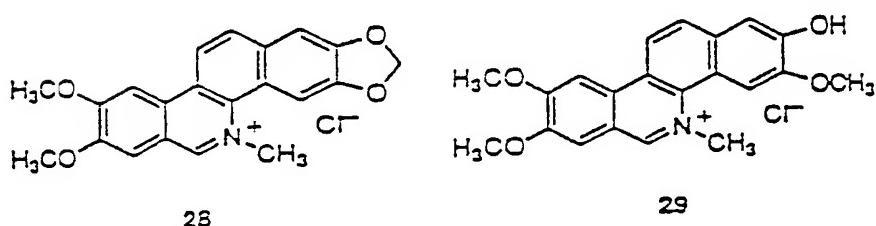


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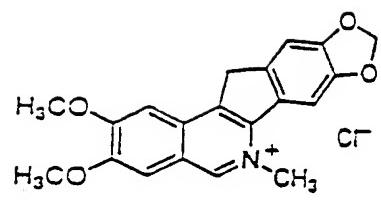
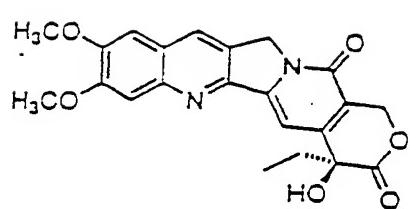


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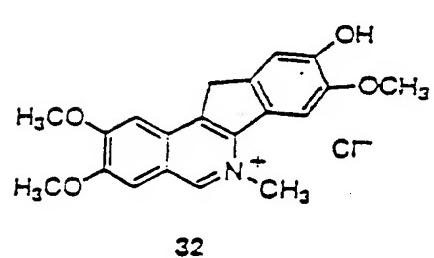
Scheme 4.

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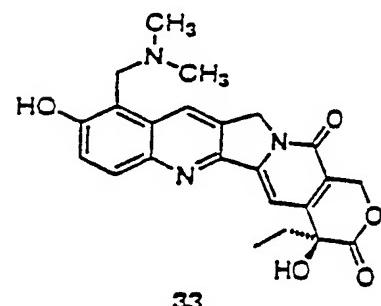
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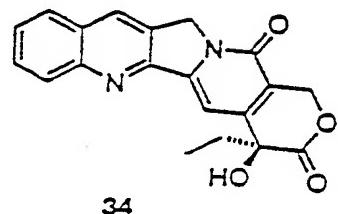
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Scheme 5.

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Dehydration, as well as dehydrogenation, of the alcohol **25** occurred in the presence of palladium in charcoal in refluxing acetic acid. Treatment of the product with aqueous NaCl provided the indenoisoquinolinium salt **27**.

Finally, we were interested in obtaining an indenoisoquinoline derivative having an acidic group which might be converted into a more water-soluble salt. The carboxylic acid **26** was obtained by oxidation of indenoisoquinoline **7** with Jones reagent.

For comparison purposes, camptothecin (**34**) and several camptothecin derivatives **33** and **30**, as well as nitidine (**28**), fagaronine (**29**), the anticancer indenoisoquinolinium species **31** and **32** (structures given in Scheme 5) were used as control agents for experiments examining topoisomerase I-mediated DNA cleavage and/or cell growth inhibition experiments.

Biological Results and Discussion

The indenoisoquinolines were examined for antiproliferative activity against the human cancer cell lines in the National Cancer Institute screen (COMPARE screening), in which the activity of each compound was evaluated with approximately 55 different cancer cell lines of diverse tumor origins. The GI₅₀ values (i.e., the concentration causing 50% growth inhibition) obtained with selected cell lines, along with the mean graph midpoint (MGM) values, are summarized in Table 1. The MGM is based on a calculation of the average GI₅₀ for all of the cell lines tested (approximately 55) in which GI₅₀ values below and above the test range (10⁻⁴ to 10⁻⁸ molar) are taken as the minimum (10⁻⁸ molar) and maximum (10⁻⁴ molar) drug concentrations used in the screening test. In addition, the relative activities of the compounds in the topoisomerase I cleavage assay are listed in Table 1. In Table 1, results of the topoisomerase I cleavage assay are listed as follows:

1) “++” designates those compounds having greater than 50% of the activity of 1μM camptothecin; 2) “+” designates those compounds having between 20% and 50% of the activity of 1μM camptothecin; 3) “±” designates those compounds having less than 20% of the activity of 1μM camptothecin; and 4) “O” designates those compounds that were inactive in the topoisomerase I cleavage assay.

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Table 1. Cytotoxicities of Indenoisoquinoline Analogs

Compd. No.	Cytotoxicity (G150 in μM) ^a								Top I Cleavage ^c
	Lung HOP-62	Colon HCT- 116	CNS SF-539	Melanoma UACC-62	Ovarian OVCAr-3	Renal SN12C	Prostate DU-145	Breast MDA- MB-435	
1	1.3	35	41	4.2	73	68	37	96	20
2	>100	>100	>100	>100	>100	>100	>100	85	++
3	--	--	--	--	--	--	--	--	0
4	--	--	--	--	--	--	--	--	0
5	61	>100	>100	84	>100	>100	>100	98	0
6	--	--	>100	>100	>100	>100	>100	81	0
7	13	3.2	4.6	4.4	74	7.4	>100	16	+
8	4.4	3.9	-	14.3	93	--	3.4	41	14
13a	22	54	-	>100	>100	>100	>100	--	45
13b	17	2.9	9.0	8.2	94	37	38	>100	30
13c	3.4	2.3	2.2	--	6.6	2.6	3.2	5.2	+
13d	16	45	--	21	78	--	16	>100	42
13e	12	>100	-	50	>100	--	4.0	>100	42
13f	20	>100	>100	>100	>100	>100	76	>100	70
13g	72	>100	>100	93	>100	88	>100	82	0

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Compd. No.	Cytotoxicity (GI50 in μM) ^a								Top I Cleavage ^c
	Lung HOP-62	Colon HCT- 116	CNS SF-539	Melanoma UACC-62	Ovarian OVCAR-3	Renal SN12C	Prostate DU-145	Breast MDA- MB-435	
13h	46	80	39	>100	58	>100	>100	>100	72
13i	>100	>100	>100	55	>100	>100	>100	>100	0
13j	24	--	>100	--	>100	>100	>100	--	95
13k	2.2	2.6	2.0	2.1	3.0	3.6	2.3	--	0
19a	0.70	1.4	--	0.99	3.7	--	0.36	10	2.4
19b	8.7	>100	--	>100	>100	--	5.1	>100	+
20	9.4	2.0	3.1	0.42	6.7	2.1	4.1	17	+
21	0.29	0.29	--	0.18	0.44	--	0.42	0.45	3.2
22	0.36	0.34	--	0.38	1.7	--	0.77	1.4	+
23	--	--	--	--	--	--	--	--	+
24	>100	>100	>100	--	>100	>100	>100	>100	48
25	30	35	50	--	24	26	41	25	48
26	87	40	--	73	>100	27	48	>100	100
27	22	13	5.6	--	1.9	2.7	58	14	13

In general, most of the new indenoisoquinolines were even less cytotoxic in human cancer cell cultures than the moderately active (MGM 20 μ M) lead compound **1**. However, a few members of the series proved to be more cytotoxic
5 than **1**, including the *N*-allyl analog **13c** (MGM 4.2 μ M), the *N*-ethyl homolog **13k** (MGM 2.4 μ M), analog **19a** (MGM 3.2 μ M) having an *N*-(4'-hydroxybutyl) substituent, and the three dihydro derivatives **20** (MGM 5.0 μ M), **21** (MGM. 0.81 μ M), and **22** (MGM 0.98 μ M). The *N*-ethyliisoquinolinium species **27** (MGM 13 μ M) and the relatively simple indenoisoquinolines **7** (MGM 16 μ M) and **8** (MGM 14 μ M),
10 both lacking substituents on the aromatic rings, were slightly more cytotoxic than **1**. Whereas the isoquinolinium salt **27** was comparable to **1** regarding topoisomerase I cleavage activity, the other more cytotoxic analogs were significantly less potent than **1** in the topoisomerase I cleavage assay.

The most potent of the new indenoisoquinolines vs. topoisomerase I
15 proved to be **26** and **27** (Table 1). Both of these compounds were examined for induction of DNA cleavage in the 3'-end-labeled PvuII/HindIII fragment of pBluescript SK(-) phagemid DNA in the presence of topoisomerase I. The results were compared with those for the lead compound **1** and camptothecin (34). Some of the cleavage sites detected in the presence of **26**, **27**, and **1** were different from those
20 induced by camptothecin (34). The indenoisoquinolines **26**, **27**, and **1** induced several topoisomerase cleavage sites that were not observed with camptothecin (34).

A wider array of compounds were tested at various concentrations and the topoisomerase inhibition data are summarized in Table 1. In general, except for **13k**, which had very weak activity, the indenoisoquinolines induced similar cleavage
25 patterns. With some of the compounds (e.g. **27**), the activity seemed to increase initially as the concentration was increased, but then it declined at higher concentrations. This is reflected in Figure 1, which was obtained after a more extensive investigation of the most potent indenoisoquinolines. The increase and following decrease in activity vs. concentration indicates that these compounds
30 suppress topoisomerase-mediated DNA cleavage at higher drug concentrations, a

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result which is similar to the bell shaped curves seen with DNA unwinding or intercalating poisons. In order to investigate the possibility that some of the most potent indenoisoquinolines could be unwinding DNA and thus causing inhibition of topoisomerase activity at higher drug concentrations, they were examined for DNA unwinding activity. The unwinding assay using supercoiled DNA in the presence of topoisomerase I is a simple procedure to detect DNA intercalation. Our results show that the indenoisoquinoline **27** in fact does unwind DNA, as does **26** at higher concentrations. On the other hand, the indenoisoquinoline **19a**, like the lead compound **1**, does not appear to unwind DNA.

Camptothecin (**34**) induces DNA strand breaks by stabilizing the cleavage complexes and inhibiting DNA re-ligation. However, increasing salt concentration can reverse the camptothecin-induced cleavage complexes, and this method has been used to compare the molecular interactions between camptothecin derivatives and topoisomerase I cleavage complexes. The cleavage sites induced by camptothecin and the indenoisoquinoline derivatives **1**, **13c**, **19a**, **26**, and **27** were reversed by salt treatment. This reversibility is consistent with the reversible trapping of topoisomerase cleavage complexes by the indenoisoquinolines.

In general, a planar indenoisoquinoline system appears to be a necessary, although not sufficient, condition for potent activity in the topoisomerase I cleavage assay. The non-planar systems **20-25** were all inactive or displayed weak activity vs. topoisomerase I (Table 1). A direct comparison can be made between the planar indenoisoquinoline **1** and the corresponding non-planar, cis dihydro compound **20**. Compound **1** displays good activity in the topoisomerase I cleavage assay, whereas the activity of **20** is weak. On the other hand, indenoisoquinolines **3-6** and **13f-13j** are all planar ring systems that are inactive as topoisomerase I inhibitors.

It is of interest to compare the results obtained with the *N*-(4'-hydroxybutyl) compound **7** with the corresponding acid **26** in the topoisomerase I cleavage assay. Both of these simple indenoisoquinolines lack substituents in the aromatic rings and differ only in the oxidation state of the terminal carbon of the *N*-

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substituent. There is a significant increase in topoisomerase I inhibitory activity in going from the alcohol **7** to the corresponding carboxylic acid **26**.

Table 2 shows the Pearson correlation coefficients derived from the GI₅₀ values for compound **1**, camptothecin (**34**) and several camptothecin derivatives **5** **33** and **30**, as well as nitidine (**28**), fagaronine (**29**), the anticancer indenoisoquinolinium species **31** and **32**, and several of the new indenoisoquinoline derivatives. The Pearson correlation coefficients quantify the degree of similarity in the cytotoxicity profiles of the compounds listed in the NCI panel of approximately 10 55 cancer cell lines. The analysis was performed using the COMPARE algorithm, which was developed to facilitate the rapid selection of compounds with similar or novel cytotoxicity profiles relative to established anticancer agents with known mechanisms of action. If the data pattern of an agent of interest correlates well with the data pattern of a known agent with a known mechanism of action, then the hypothesis is formed that the agent of interest may have the same mechanism of 15 action as that of the known agent. In the present case, the dihydroindenoisoquinoline derivative **20** correlates well with the camptothecins **30**, **33**, and **34**, suggesting that the cytotoxicity of **20** may be due to its topoisomerase I inhibitory activity.

Table 2. Pearson Correlations Derived From GI₅₀ Values.

Cpd	28	29	1	30	31	20	32	23	33	13k	34
28	1.00	0.59	0.36	0.29	0.26	0.16	0.25	0.32	0.25	0.03	0.30
29	0.59	1.00	0.47	0.54	0.23	0.39	0.61	0.48	0.49	-0.01	0.48
1	0.36	0.47	1.00	0.73	0.28	0.59	0.58	0.75	0.58	0.23	0.56
30	0.29	0.54	0.73	1.00	0.41	0.74	0.73	0.79	0.83	0.17	0.78
31	0.26	0.23	0.28	0.41	1.00	0.39	0.57	0.53	0.24	0.55	0.23
20	0.16	0.40	0.59	0.74	0.39	1.00	0.72	0.73	0.69	0.25	0.73
32	0.25	0.61	0.58	0.73	0.57	0.72	1.00	0.77	0.72	0.25	0.68
23	0.32	0.48	0.75	0.79	0.53	0.73	0.77	1.00	0.67	0.21	0.64
33	0.25	0.56	0.58	0.83	0.24	0.69	0.72	0.67	1.00	0.18	0.87
13k	0.03	-0.01	0.23	0.17	0.55	0.25	0.25	0.21	0.18	1.00	0.14

34	0.30	0.48	0.56	0.78	0.23	0.73	0.68	0.64	0.87	0.14	1.00
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Since a number of topoisomerase I poisons also inhibit topoisomerase II, we tested the induction of topoisomerase II cleavage complexes by indenoisoquinolines. Our results show that compound **26** induced topoisomerase II cleavage complexes at sites which often did not overlap with the topoisomerase II sites induced by VP-16 (etoposide). Compound **27** had only marginal topoisomerase II activity at 100 µM and compounds **13c**, **19a** and **1** had no effect on topoisomerase II cleavage activity. Compounds **7** and **8** also exhibited weak topoisomerase II activity and compounds **13b**, **13k**, **20**, **21** and **22** had no effect on topoisomerase II cleavage. These results indicate that the indenoisoquinolines are prominent topoisomerase I inhibitors, except for the two derivatives **26** and **27** that also produce DNA unwinding.

The objective of maximizing the cytotoxicity of indenoisoquinoline compounds against tumor or cancer cell lines was realized in the indenoisoquinolines **7**, **8**, **13c**, **13k**, **19a**, **20**, **21**, **22**, and **27**, all of which displayed a lower MGM than the lead compound **1** (Table 1). Further, several topoisomerase I inhibitors were synthesized which rival the topoisomerase activity of **1**, including **13c**, **19a**, **26**, and **27**. One obvious point of further interest is that with the possible exception of **19a**, the two activities did not maximize in the same compounds, suggesting that the activity of some of the more cytotoxic compounds may not be due to their activity vs. topoisomerase. The situation is complicated by such factors as cellular uptake and possible conversion of parent compounds to metabolites which may have increased activity vs. topoisomerase I.

25 Examples

The following examples demonstrate the syntheses of several embodiments of the compounds of the present invention. Melting points were determined in capillary tubes and are uncorrected. Infrared spectra were obtained 30 using CHCl₃ as the solvent unless otherwise specified. ¹H NMR spectra were

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obtained using CDCl_3 as solvent and TMS as internal standard. ^1H NMR spectra were determined at 300 MHz. Chemical ionization mass spectra (CIMS) were determined using isobutane as the reagent gas. Microanalyses were performed at the Purdue University Microanalysis Laboratory. Analytical thin-layer chromatography was 5 carried out on Analtech silica gel GF 1000 micron glass plates. Compounds were visualized with short wavelength UV light or phosphomolybdic acid indicator. Silica gel flash chromatography was performed using 230-400 mesh silica gel.

Example 1: 6-Ethyl-5,6-dihydro-5,11-diketo-11*H*-indeno[1,2-*c*]isoquinoline (3): Ethylamine (0.2 mL, 3 mmol) was added to a stirred solution of 10 benz[*d*]indeno[1,2-*b*]pyran-5, 11-dione (**2**) (0.49 g, 2 mmol) in CHCl_3 (10 mL). The bright orange mixture stirred overnight. To the reaction mixture CHCl_3 (100 mL) was added and the mixture washed with H_2O (3 x 25 mL) and brine (1 x 25 mL), dried (MgSO_4), and concentrated under reduced pressure to give an orange-red solid (0.43 g, 75%): mp 188-189°C; IR (thin film) 2986, 1690, 1656, 1611, 1549, 1503, 1430, 15 1320, 1197, 991 cm^{-1} ; ^1H NMR (DMSO-d_6 , 300 MHz) δ 8.66 (d, 1 H, J = 8.3 Hz), 8.32 (d, 1 H, J = 7.9 Hz), 7.69 (dt, 1 H, J = 8.4, 1.4 Hz), 7.60 (dd, 1 H, J = 8.0, 1.4 Hz), 7.52 (d, 1 H, J = 6.9 Hz), 7.40 (m, 3 H), 4.56 (q, 2 H, J = 7.2 Hz), 1.53 (t, 3 H, J = 7.2 Hz); CIMS, m/z (relative intensity) 276 (MH^+ , 100). Anal. Calcd for $\text{C}_{18}\text{H}_{13}\text{NO}_2$: C, H, N.

Example 2: 5,6-Dihydro-5,11-diketo-6-propyl-11*H*-indeno[1,2-*c*]isoquinoline (4). Propylamine (0.3 mL, 3 mmol) was added to a stirred solution of benz[*d*]indeno[1,2-*b*]pyran-5, 11-dione (**2**) (0.49 g mmol) in CHCl_3 (10 mL). The red solution stirred overnight before CHCl_3 (75 mL) was added and the mixture washed with H_2O (3 x 20 mL) and brine (1 x 20 mL), dried (MgSO_4), and concentrated under 25 reduced pressure to give a yellow-orange solid (0.32 g, 55%): mp 166-167°C; IR (neat) 2967, 1660, 1502, 1427, 1317, 1193, 959 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz) δ 8.69 (d, 1 H, J = 8.0 Hz), 8.33 (d, 1 H, J = 9.0 Hz), 7.70 (td, 1 H, J = 9.0, 3.0 Hz), 7.62 (d, 1 H, J = 6.2 Hz), 7.40 (m, 4 H), 4.46 (t, 2 H, J = 8.0 Hz), 1.92 (m, 2 H), 1.12 (t, 3 H, J = 7.4 Hz); CIMS m/z (relative intensity) 290 (MH^+ , 100). Anal. Calcd for 30 $\text{C}_{19}\text{H}_{15}\text{NO}_2$: C, H, N.

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Example 3: 6-Cyclopropyl-5,6-dihydro-5,11-diketo-11*H*-

indeno[1,2-*c*]isoquinoline (5). Cyclopropylamine (10 mL) was added to a stirred solution of benz[*d*]indeno[1,2-*b*]pyran-5,11-dione (2) (0.28 g, 1.1 mmol) in CHCl₃ (10 mL). The red solution stirred overnight before CHCl₃ (50 mL) was added and the mixture washed with H₂O (3 x 20 mL) and brine (1 x 20 mL), dried (MgSO₄), and concentrated under reduced pressure to give a red solid (0.3 g, 91%): mp 206-208°C; IR (thin film) 3751, 1665, 1500, 1420, 1311, 1083, 950 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.62 (d, 1 H, *J* = 7.7 Hz), 8.29 (d, 1 H, *J* = 8.4 Hz), 7.88 (d, 1 H, *J* = 7.0 Hz), 7.69 (td, 1 H, *J* = 6.9, 1.2 Hz), 7.59 (dd, 1 H, *J* = 6.1, 1.3 Hz), 7.40 (m, 3 H), 3.37 (m, 1 H), 1.45 (q, 2 H, *J* = 6.8 Hz), 0.99 (m, 2 H); CIMS *m/z* (relative intensity) 288 (MH⁺, 100). Anal. Calcd for C₁₉H₁₃NO₂: C, H, N.

Example 4: 5,6-Dihydro-5,11-diketo-6-(methoxycarbonylmethyl)-

11*H*-indeno[1,2-*c*]isoquinoline (6): Triethylamine (2.7 mL, 19.4 mmol) was added to a stirred solution of glycine methyl ester hydrochloride (1.57 g, 12.5 mmol) in chloroform (30 mL). After 1 h, benz[*d*]indeno[1,2-*b*]pyran-5, 11-dione (2) (1.24 g, 5.0 mmol) was added to the mixture. The red mixture stirred an additional 4 h before CHCl₃ (100 mL) was added and the mixture washed with H₂O (3 x 50 mL) and brine (1 x 50 mL), dried (MgSO₄), and concentrated under reduced pressure to give an orange-red solid (1.48 g, 92%): mp 248-251°C; IR (thin film) 2956, 1735, 1667, 1609, 1502, 1426, 1227, 981 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.68 (d, 1 H, *J* = 8.0 Hz), 8.32 (d, 1 H, *J* = 8.2 Hz), 7.73 (td, 1 H, *J* = 7.1, 1.3 Hz), 7.61 (m, 1 H), 7.47 (td, 1 H, *J* = 7.1, 1.1 Hz), 7.37 (m, 2 H), 7.26 (m, 1 H), 5.34 (s, 2 H), 3.79 (s, 3 H); CIMS *m/z* (relative intensity) 320 (MH⁺, 100). Anal. Calcd for C₁₉H₁₃NO₄: C, H, N.

Example 5: 5,6-Dihydro-6-(4-hydroxy-1-butyl)-5,11-diketo-11*H*-

indeno[1,2-*c*]isoquinoline (7): 4-Amino-1-butanol (0.891 g, 10 mmol) was added to a chloroform (30 mL) solution of benz[*d*]indeno[1,2-*b*]pyran-5, 11-dione (2) (2.48 g, 10 mmol) and the reaction mixture was stirred at room temperature 2 days. The reaction mixture turned dark red. The reaction mixture was taken in chloroform (100 mL) and washed with water (2 x 50 mL), 0.5 N HCl (50 mL), brine (100 mL) and dried (Na₂SO₄) and

concentrated to give the crude product. The product was filtered through a short column of silica gel and the polar fraction concentrated to afford a reddish brown solid which was crystallized from isopropanol to yield the product (2.56 g, 80%): mp 160 - 162 °C; IR (KBr) 3300, 1695, 1645, 1615 cm⁻¹; ¹H NMR (CDCl₃) δ 8.63 (d, *J* = 8.1 Hz, 1 H), 8.26 (d, *J* = 8.1 Hz, 1 H), 7.70 - 7.15 (m, 6 H), 4.51 (t, *J* = 7.8 Hz, 2 H), 3.77 (t, *J* = 6.1 Hz, 2 H), 1.99 (p, *J* = 8.0 and 7.5 Hz, 2 H), 1.83 (s, 1 H, D₂O exchangeable). Anal. Calcd for C₂₀H₁₇NO₃: C, H, N.

5 **Example 6: 5,6-Dihydroxy-6-(5-hydroxy-1-pentyl)-5,11-diketo-11*H*-indeno[1,2-*c*]isoquinoline (8).** 5-Amino-1-pentanol (1.03 g, 10 mmol) was added to a chloroform (20 mL) solution of benz[*d*]indeno[1,2-*b*]pyran-5, 11-dione (**2**) (2.48 g, 10 mmol) and the reaction mixture was stirred at room temperature overnight. The reaction mixture turned dark red. The reaction mixture was taken in chloroform (100 mL) and washed with water (2 x 50 mL), 0.5 N HCl (50 mL), brine (100 mL) and dried (Na₂SO₄) and concentrated to give the crude product. The TLC showed traces of starting material. The product was filtered through a short column of silica gel and the polar fraction concentrated to get a reddish brown solid which was crystallized from isopropanol to afford the product (2.53 g, 76%): mp 146-148 °C; IR (KBr) 2996, 1698, 1642, 1615 cm⁻¹; ¹H NMR (CDCl₃) δ 8.63 (d, *J* = 8.1 Hz, 1 H), 8.27 (d, *J* = 8.1 Hz, 1 H), 7.67 (d, *J* = 8.4 Hz, 1 H), 7.56 (d, *J* = 6.8 Hz, 1 H), 7.45 - 15 7.30 (m, 4 H), 4.47 (t, *J* = 7.9 Hz, 2 H), 3.71 (t, *J* = 5.9 Hz, 2 H), 1.92 (p, *J* = 7.9 and 7.4 Hz, 2 H), 1.82 (s, 1 H, D₂O exchangeable), 1.78 - 1.55 (m, 4 H); CIMS *m/z* (relative intensity) 334 (MH⁺, 100). Anal. Calcd for C₂₁H₁₉NO₃: C, H, N.

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25 **Example 7: cis-4-Carboxy-3,4-dihydro-N-methyl-3-(3',4'-methylenedioxyphenyl)-1(2*H*)isoquinolone (12a):** Homophthalic anhydride (**10a**) (0.81 g, 5 mmol) was added to a stirred solution of 3,4-methylenedioxybenzylidenemethylamine (**11a**) (0.82 g, 5 mmol) in chloroform (5 mL). After 30 min, the precipitated product was filtered from the yellow solution and washed with chloroform to give a pale yellow solid (1-2 g, 74%): mp 165-167 °C; ¹H NMR (DMSO-d₆) δ 7.99 (d, *J* = 7.5 Hz, 1 H), 7.48 (m, 3 H), 6.76 (d, *J* = 8.0 Hz, 1 H),

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6.52 (d, $J = 8.0$ Hz, 1 H), 6.43 (s, 1 H), 5.93 (s, 2 H), 5.03 (d, $J = 6.2$ Hz, 1 H), 4.64 (d, $J = 6.1$ Hz, 1 H), 2.89 (s, 3 H); CIMS m/z (relative intensity) 326 (MH^+ , 100).

Example 8: 5,6-Dihydro-5,11-diketo-6-methyl-8,9-methylenedioxy-11H-indeno[1,2-c]isoquinoline (13a)

5 Thionyl chloride (8.1 mL) was added with stirring to the cis acid **12a** (0.7 g, 2.1 mmol). The yellowish-brown mixture became orange within 15 min and after 30 min was red. After 4 h, the reaction mixture was diluted with benzene (25 mL) and evaporated to dryness. The brownish-red solid was recrystallized from methanol and passed through a short column (SiO_2) and eluted with chloroform to give a brown solid (0.14 g, 24%): mp 10 310-312 °C; IR (thin film) 2358, 1652, 1540, 1506, 1292 cm^{-1} ; ^1H NMR (DMSO-d_6) δ 8.43 (d, $J = 8.0$ Hz, 1 H), 8.16 (d, $J = 8.0$ Hz, 1 H), 7.75 (t, $J = 7.5$ Hz, 1 H), 7.56 (s, 1 H), 7.44 (t, $J = 7.6$ Hz, 1 H), 7.15 (s, 1 H), 6.19 (s, 2 H), 3.92 (s, 3 H); CIMS m/z (relative intensity) 306 (MH^+ , 100). Anal. Calcd for $\text{C}_{18}\text{H}_{11}\text{NO}_4$: C, H, N.

Example 9: 3,4-Methylenedioxybenzylidenebutylamine (11b)

15 Piperonal (7.5 g, 50 mmol) and *n*-butylamine (6 mL, 75 mmol) were stirred in chloroform (100 mL) in the presence of anhydrous MgSO_4 (5 g) at room temperature for 4 h. The mixture was filtered and the residue was washed with chloroform (20 mL). The combined filtrate was concentrated under reduced pressure to afford a yellow oil (9.8 g, 96%): IR (neat) 1649, 1643, 1604 cm^{-1} ; ^1H NMR (CDCl_3) δ 8.11 (s, 1 H), 7.31 (d, $J = 1.2$ Hz, 1 H), 7.06 (dd, $J = 1.2$ and 7.9 Hz, 1 H), 6.79 (d, $J = 7.8$ Hz, 1 H), 5.95 (s, 2 H), 3.53 (t, $J = 6.6$ Hz, 2 H), 1.63 (p, $J = 7.3$ Hz, 2 H), 1.37 (heptet, $J = 7.3$ Hz, 2 H), 0.91 (t, $J = 7.3$ Hz, 3 H).

Example 10: cis-N-(1-Butyl)-4-carboxy-3,4-dihydro-3-(3',4'-methylenedioxyphenyl)-1(2H)-isoquinolone (12b): Homophthalic anhydride (**10a**) (3.24 g, 20 mmol) was added to a chloroform (20 mL) solution of the imine **11b** (4.1 g, 20 mmol) and the mixture was stirred at room temperature for 45 min, after which the TLC showed the complete disappearance of the starting materials. The reaction mixture was concentrated to remove chloroform completely. The residue was dissolved in hot ethyl acetate (100 mL) and left at room temperature for 12 h. The 20 colorless crystals that separated were filtered and dried to give pure **12b** (6.57 g, 30

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89%): mp 178-181 °C; IR (KBr) 1712, 1634, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ 8.08 (dd, J = 1.0 and 7.5 Hz, 1 H), 7.52 (d, J = 7.5 Hz, 1 H), 7.40-7.28 (m, 2 H), 6.51-6.45 (m, 2 H), 6.37 (s, 1 H), 5.75 (dd, J = 1.1 and 6.4 Hz, 2 H), 4.87 (d, J = 6.2 Hz, 1 H), 4.51 (d, J = 6.2 Hz, 1 H), 3.93 (dt, J = 7.2 and 6.6 Hz, 1 H), 2.73 (dt, J = 7.2 and 6.6 Hz, 1 H), 1.52 (p, J = 7.2 Hz, 2 H), 1.26 (heptet, J = 7.3) Hz, 2 H), 0.83 (t, J = 7.2 Hz, 3 H); CIMS m/z (relative intensity) 368 (MH⁺, 100); EIMS m/z (relative intensity) 367 (M⁺, 5), 322 (30), 135 (100). Anal. Calcd for C₂₁H₂₁NO₅: C, H, N.

Example 11: **6-(1 -Butyl)-5,6-dihydro-5,11-diketo-8,9-methylenedioxy-11H-indeno[1,2 -c]isoquinoline (13b):** Thionyl chloride (30 mL) was added dropwise to the acid **12b** (3.35 g, 0.089 ml) with stirring. The resulting solution was stirred at room temperature for 12 h, after which the solution turned dark pink. Benzene (20 mL) was added to the reaction mixture and it was concentrated under reduced pressure. The resulting residue was purified by column chromatography (acetone:hexane, 1:4) followed by crystallization (EtOAc/Hexane) to obtain pure indenoisoquinoline **13b** (1.37 g, 44%): mp 200-201 °C; IR (KBr) 1691, 1665, 1631 cm⁻¹; ¹H NMR (CDCl₃) δ 8.50 (d, J = 8.1 Hz, 1 H), 8.21 (d, J = 8 Hz, 1 H), 7.61 (t, J = 8 Hz, 1 H), 7.34 (t, J = 8 Hz, 1 H), 6.98 (s, 1 H), 6.87 (s, 1 H), 6.03 (s, 2 H), 4.34 (t, J = 8 Hz, 2 H), 1.80 (p, J = 8 Hz, 2 H), 1.51 (heptet, J = 8 Hz, 2 H), 1.01 (t, J = 8 Hz, 3 Hz); ¹³C NMR (CDCl₃) δ 189.01, 163.1, 154.85, 151.21, 148.97, 133.58, 132.18, 132.05, 130.50, 128.29, 126.39, 122.82, 122.69, 107.48, 105.12, 104.84, 102.57, 44.13, 31.33, 20.1, 13.73; CIMS m/z (relative intensity) 348 (MH⁺, 100); EIMS m/z (relative intensity) 347 (M⁺, 60), 330 (10), 318 (30), 291 (100). Anal. Calcd for C₂₁H₁₇NO₄: C, H, N.

Example 12: **3,4-Dimethoxybenzylideneallylamine (11c):** Allylamine (6 mL, 80 mmol) was added to a solution of 3,4-dimethoxybenzaldehyde (8.3 g, 50 mmol) in dichloromethane (50 mL) in the presence of anhydrous magnesium sulfate (5 g) and the reaction mixture was stirred at room temperature overnight. The reaction mixture was filtered, the residue washed with chloroform (10 mL) and the combined filtrate was concentrated under reduced pressure to afford **11** as a yellow oil (10.18 g, 99%): IR (neat) 1692, 1679, 1646, 1604 cm⁻¹; ¹H NMR

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(CDCl₃) δ 8.14 (s, 1 H), 7.40 (d, *J* = 1.8 Hz, 1 H), 7.11 (dd, *J* = 1.8 and 8 Hz, 1 H), 6.82 (d, *J* = 8 Hz, 1 H), 6.10-5.90 (m, 1 H), 5.20 (dd, *J* = 1.8 and 17.4 Hz, 1 H), 5.10 (dd, *J* = 1.3 and 10 Hz, 1 H), 4.20 (d, *J* = 6.3 Hz, 2 H), 3.88 (s, 3 H), 3.85 (s, 3 H).

Example 13: *cis-N-Allyl-4-carboxy-3,4-dihydro-6,7-dimethoxy-3-*

5 (*3',4'-methylenedioxy phenyl*)-1(2*H*)isoquinolone (**12c**). 4,5-Dimethoxyhomophthalic anhydride (**10b**) (1.11 g, 5 mmol) was added to a chloroform (10 mL) solution of the imine **11c** (0.945 g, 5 mmol) and the mixture was stirred at room temperature for 45 min, after which the TLC showed the complete disappearance of the starting materials and a white precipitate formed in the reaction mixture. The precipitated product was filtered off and washed with chloroform (5 mL) and dried to give pure **12c** (1.43 g, 70%): mp 235-238 °C; IR (KBr) 3000, 1736, 1686, 1615 cm⁻¹; ¹H NMR (DMSO-d₆) δ 13.0 (bs, 1 H), 7.52 (s, 1 H), 7.13 (s, 1 H), 6.76 (d, *J* = 8.5 Hz, 1 H), 6.52 (d, *J* = 8.5 Hz, 1 H), 6.44 (s, 1 H), 5.94 (s, 2 H), 5.85-5.70 (m, 1 H), 5.16 (dd, *J* = 3.5 & 17.5 Hz, 2 H), 4.92 (d, *J* = 6.5 Hz, 1 H), 4.57 (d, *J* = 6.5 Hz, 1 H), 3.82 (s, 3 H), 3.75 (s, 3 H), 3.20 - 3.10 (m, 2 H). CIMS *m/z* (relative intensity) 412 (MH⁺, 100). Anal. Calcd for C₂₂H₂₁NO₇: C, H, N.

Example 14: *6-Allyl-2,3-dimethoxy-5,6-dihydro-5,11-oxo-8,9-(methylenedioxy)-11*H*-indeno[1,2-c]isoquinoline (**13c**)*:

Treatment of **12c** (2.05 g, 5 mmol) with Eaton's reagent (10% P₂O₅, in methanesulfonic acid, 60 mL) at room temperature with stirring in an open flask for 24 h resulted in a mixture of **22** and **13c**. The two products were separated by column chromatography on silica gel (230-400 mesh) using hexane:acetone (4:1) to afford **22** (842 mg, 43%) and **12c** as a purple solid product (588 mg, 30%) after recrystallization from ethyl acetate:hexane: mp 290-294 °C; IR (KBr) 2370, 1698, 1653, 1551, 1484; ¹H NMR (CDCl₃) δ 7.97 (s, 1 H), 7.63 (s, 1 H), 6.99 (s, 1 H), 6.91 (s, 1 H), 6.20-6.05 (m, 1 H), 6.06 (s, 2 H), 5.31 (d, *J* = 10.5 Hz, 1 H), 5.20 - 5.00 (m, 3 H), 3.33 (s, 3 H), 3.97 (s, 3 H). Anal. Calcd for C₂₂H₁₇NO₆: C, 67.52; H, 4.38; N, 3.58. Found: C, 67.18; H, 4.32; N, 3.31.

Example 15: *cis-N-(1-Butyl)-4-carboxy-3,4-dihydro-6,7-*

dimethoxy-3-(*3',4'-methylenedioxyphenyl*)-1(2*H*)isoquinolone (**12d**). 4,5-Dimethoxyhomophthalic anhydride **10b** (2.22 g, 10 mmol) was added to a chloroform

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(10 mL) solution of the imine (**11b**) (2.1 g, 10 mmol) and the mixture was stirred at room temperature for 45 min, after which the TLC showed the complete disappearance of the starting materials and a white precipitate formed in the reaction mixture. The precipitated product was filtered off and washed with chloroform (5 mL) and dried to give pure **12d** (3.45 g, 81%): mp 242-244°C; IR (KBr) 1732, 1640, 1610, 1600 cm⁻¹; ¹H NMR (CDCl₃ + DMSO-d₆) δ 7.56 (s, 1 H), 7.08 (s, 1 H), 6.55 - 6.48 (m, 2 H), 6.40 (s, 1 H), 5.79 (d, J = 2.5 Hz, 2 H), 4.86 (d, J = 6.2 Hz, 1 H), 4.45 (d, J = 6.2 Hz, 1 H), 3.88 (dt, J = 7.4 and 6.1 Hz, 1 H), 2.71 (dt, J = 7.5 and 6.1 Hz, 1 H), 1.49 (p, J = 7.3 Hz, 2 H), 1.26 (heptet, J = 7.3 Hz, 2 H), 0.83 (t, J = 7.3 Hz, 3 H).
10 Anal. Calcd for C₂₃H₂₅NO₇: C, H, N.

Example 16: **6-(1-Butyl)-5,6-dihydro-5,11-diketo-2,3-dimethoxy-8,9-methylenedioxy11H-indeno[1,2-c]isoquinoline (13d).** Thionyl chloride (30 mL) was added dropwise to the acid **12d** (2.135 g, 5 mmol) with stirring. The resulting solution was stirred at room temperature for 12 h after which the solution turned dark pink. Benzene (20 mL) was added to the reaction mixture and it was concentrated under reduced pressure. Benzene (50 mL) was added to the resulting residue and the pink solid was filtered off to obtain pure indenoisoquinoline **13d** (1.3 g, 65%): mp 280-284°C; IR (KBr) 1699, 1653, 1646, 1578 cm⁻¹; ¹H NMR (CDCl₃) δ 7.99 (s, 1 H), 7.62 (s, 1 H), 7.04 (s, 1 H), 6.92 (s, 1 H), 6.07 (s, 2 H), 4.39 (t, J = 7.6 Hz, 2 H), 4.01 (s, 3 H), 3.96 (s, 3 H), 1.82 (p, J = 7.3 Hz, 2 H), 1.68 - 1.55 (m, 2 H), 1.02 (t, J = 7.3 Hz, 3 Hz). Anal. Calcd for C₂₃H₂₁NO₆ 0.1 H₂O: C, H, N.
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Example 17: **3,4-Methylenedioxybenzylidenebenzylamine (11e).** Piperonal (4.5 g, 30 mmol) and benzylamine (3.21 g, 30 mmol) were stirred in methylene chloride (30 mL) in the presence of anhydrous MgSO₄ (5 g) at room temperature for 4 h. The mixture was filtered and the residue was washed with methylene chloride (20 mL) and the combined filtrate was concentrated under reduced pressure to afford a white solid (7.03 g, 98 %): mp 69-70°C; IR (KBr) 1638, 1618, 1602 cm⁻¹; ¹H NMR (CDCl₃) δ 8.18 (s, 1 H), 7.33 (d, J = 1.3 Hz, 1 H), 7.30 - 7.10 (m, 5 H), 7.06 (dd, J = 1.3 and 8.0 Hz, 1 H), 6.74 (d, J = 8 Hz, 1 H), 5.90 (s, 2 H), 4.69 (s, 2 H).
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Example 18: cis-N-Benzyl-4-carboxy-3,4-dihydro-6,7-dimethoxy-

3-(3',4'-methylenedioxyphenyl)-1(2H)isoquinolone (12e): 4,5-

Dimethoxyhomophthalic anhydride (**10b**) (1.11 g, 5 mmol) was added to a chloroform (10 mL) solution of the imine **11e** (1.19 g, 5 mmol) and the mixture was stirred at room temperature for 2 h, after which the TLC showed the complete disappearance of the starting materials and a white precipitate formed in the reaction mixture. The precipitated product was filtered off and washed with chloroform (5 mL) and dried to give pure **12e** (1.89 g, 82%): mp 262-264°C; IR (KBr) 1736, 1654, 1647, 1618, 1595, 1575 cm⁻¹; ¹H NMR (DMSO-d₆) δ 7.56 (s, 1 H), 7.35 - 7.20 (m, 5 H), 7.13 (s, 1 H), 6.75 (d, J = 8.3 Hz, 1 H), 6.51 (d, J = 8.1 Hz, 1 H), 6.43 (s, 1 H), 5.93 (s, 2 H), 5.25 (d, J = 15.6 Hz, 1 H), 4.86 (d, J = 5.6 Hz, 1 H), 4.51 (d, J = 5.3 Hz, 1 H), 3.83 (s, 3 H), 3.74 (s, 3 H), 3.39 (d, J = 15.6 Hz, 1 H).

Example 19: 6-Benzyl-5,6-dihydro-5,11-diketo 2,3-dimethoxy-8,9-

methylenedioxy-11H-indeno[1,2-c]isoquinoline (13e). Thionyl chloride (10 mL)

was added dropwise to the acid **12e** (1.15 g, 1.5 mmol) with stirring. The resulting mixture was stirred at room temperature for 5 h, after which the solution turned purple. Benzene (20 mL) was added to the reaction mixture and it was concentrated under reduced pressure. Carbon tetrachloride was added to the resulting residue and the undissolved solid was filtered off to obtain pure indenoisoquinoline **13e** (0.716 g, 65%): mp 310-312°C; IR (KBr) 1695, 1652, 1619, 1578 cm⁻¹; ¹H NMR (CDCl₃) δ 8.02 (s, 1 H), 7.66 (s, 1 H), 7.4 - 7.20 (m, 5 H), 7.02 (s, 1 H), 6.74 (s, 1 H), 5.99 (s, 2 H), 5.69 (s, 2 H), 4.04 (s, 3 H), 3.97 (s, 3 H); ¹³C NMR (CDCl₃) δ 162.54, 155.03, 148.72, 135.44, 132.52, 130.22, 129.19, 127.67, 125.64, 108.32, 105.24, 103.03, 102.47, 56.31, 47.80 and 56.03. Anal. Calcd for C₂₆H₁₉NO₆ 0.8 H₂O: C, H, N.

Example 20: 3,4-Methylenedioxybenzylidene-p-anisidine (11f).

Piperonal (15 g, 0.1 mol) and *p*-anisidine (12.3 0.1 mol) were stirred in methylene chloride (100 mL) in the presence of anhydrous MgSO₄, (5 g) at room temperature for 4 h. The mixture was filtered, the residue was washed with methylene chloride (20 mL), and the combined filtrate was concentrated under reduced pressure to afford a yellow solid. The crude product was crystallized in 95% ethanol to give white

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crystalline solid (22.38 g, 87 %): mp 113-114°C; IR (KBr) 1636 and 1617 cm⁻¹; ¹H NMR (CDCl₃) δ 8.33 (s, 1 H), 7.51 (d, *J* = 1.2 Hz, 1 H), 7.25 - 7.10 (m, s), 6.95-6.80 (m, 3 H), 6.01 (s, 2 H), 3.80 (s, 3 H).

Example 21: *cis-N-(p-Anisyl)-4-carboxy-3,4-dihydro-6,7-*

5 **dimethoxy-3-(3',4'-methylene-dioxyphenyl)-1(2*H*)isoquinolone (12f).** 4,5-Dimethoxyhomophthalic anhydride (**10b**) (1.11 g, 5 mmol) was added to a chloroform (10 mL) solution of the imine **11f** (1.275 g, 5 mmol) and the mixture was stirred at room temperature for 12 h, after which the TLC showed the complete disappearance of the starting materials and a white precipitate formed in the reaction mixture. The 10 precipitated product was filtered off, washed with chloroform (5 mL), and dried to afford pure **12f** (1.36 g, 60%): mp >350°C; IR (KBr) 1644, 1639, 1599 cm⁻¹; ¹H NMR (DMSO-d₆) δ 7.60-7.30 (m, 5 H), 7.20-6.80 (m, 4 H), 6.10 (s, 2 H), 5.30 (d, *J* = 6 Hz, 1 H), 4.77 (d, *J* = 6 Hz, 1 H), 3.78 (s, 3 H), 3.71 (s, 3 H), 3.01 (s, 3 H).

Example 22: *6-(p-Anisyl)-2,3-dimethoxy-5,6-dihydro-5,11-diketo-*

15 **8,9-methylenedioxy-11*H*-indeno[1,2-*c*]isoquinoline (13f).** Thionyl chloride (9 mL) was added dropwise to the acid **12f** (0.822 g, 2 mmol) with stirring. The resulting solution was stirred at room temperature for 5 h, after which the solution turned purple. Benzene (20 mL) was added to the reaction mixture and it was concentrated under reduced pressure. The resulting residue was passed through a short column of 20 silica gel (230-400 mesh) eluting with chloroform. Concentration of the eluent resulted in a pink solid which was crystallized from ethyl acetate to obtain pure indenoisoquinoline **13f** (0.436 g, 53%): mp 360-364°C; IR (KBr) 1692, 1652, 1625 and 1552 cm⁻¹; ¹H NMR (CDCl₃) δ 7.94 (s, 1 H), 7.60 (s, 1 H), 7.34 (d, *J* = 8.1 Hz, 2 H), 7.24 (s, 1 H), 7.10 (d, *J* = 8 Hz, 2 H), 6.88 (s, 1 H), 5.90 (s, 2 H), 5.05 (s, 1 H), 4.02 (s, 3 H), 3.93 (s, 3 H), 3.91 (s, 3 H); CIMS *m/z* (relative intensity) 458 (MH⁺, 100). Anal. Calcd for C₂₆H₁₉NO₇: C, H, N.

Example 23: *3,4-Dibenzylxybenzylidenemethylamine (11g).* 3,4-

Dibenzylxybenzaldehyde (7.96 g, 25.0 mmol) was added to a 40% aqueous solution of methylamine (10 mL) and the reaction mixture was stirred at room temperature for 30 3 h. The mixture was extracted with ether (4 x 75 mL), the ether layers were

combined, and the solution washed with saturated aqueous sodium chloride (75 mL), dried (MgSO_4), and concentrated under reduced pressure to give an off-white solid (7.7 g, 94%): mp 56-57°C; IR (KBr) 3031, 2936, 2832, 1648, 1600, 1582, 1509, 1454, 1431, 1267, 1171, 1137, 1017, 735, 696 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz) δ 8.14 (s, 1 H), 7.35 (m, 11 H), 7.11 (dd, J = 8.1, 1.0 Hz, 1 H), 6.93 (d, J = 8.1 Hz, 1 H), 5.18 (s, 4 H), 3.46 (s, 3 H); CIMS m/z (relative intensity) 332 (MH^+ , 100). Anal. Calcd for $\text{C}_{31}\text{H}_{21}\text{NO}_2$: C, H, N.

Example 24: *cis*-3-(3',4'-Dibenzylloxyphenyl)-4-carboxy-3,4-dihydro-*N*-methyl-1-2*H*-isoquinolone (12g). Homophthalic anhydride (10a) (0.81 g, 5 mmol) was added to a stirred solution of 3,4-dibenzylloxybenzylidenemethylamine (11g) (1.66 g, 5 mmol) in chloroform (5 mL). After 30 min, ether was added and the resulting precipitate was filtered and washed with ether to give a pale yellow solid (0.9 g, 36%): mp 170-172°C; IR (thin film) 3030, 1731, 1625, 1514, 1263, 1137, 1014 cm^{-1} ; ^1H NMR (CDCl_3 , 300 MHz) δ 8.19 (dd, J = 6.5, 1.9 Hz, 1 H), 7.36 (m, 10 H), 7.09 (d, J = 4.9, 1 H), 6.74 (d, J = 8.9, 1 H), 6.68 (d, J = 8.9, 1 H), 6.51 (m, 2 H), 5.03 (d, J = 7.2, 2 H), 4.92 (d, J = 6.1, 2 H), 4.8 (d, J = 6.3, 2 H), 4.5 (d, J = 6.2, 2 H), 2.98 (s, 3 H); CIMS m/z (relative intensity) 494 (MH^+ , 100). Anal. Calcd for $\text{C}_{31}\text{H}_{27}\text{NO}_5$: C, H, N.

Example 25: 8,9-Dibenzyloxy-5,6-dihydro-5,11-diketo-6-methyl-11*H*-indeno[1,2-c]isoquinoline (13g). Thionyl chloride (8.1 mL) was added with stirring to the *cis* acid 12g (0.7 g, 2.1 mmol). The result was a yellowish-brown mixture that became orange within 15 min and after 30 min was red. After 4 h, the reaction mixture was diluted with benzene (25 mL) and evaporated to dryness. The brownish-red solid was recrystallized from methanol and passed through a short column (SiO_2), eluting with chloroform, to give a brown solid (0.14 g, 24%): MP 198-200°C; ^1H NMR (DMSO-d_6) δ 8.43 (d, J = 8.0 Hz, 1 H), 8.16 (d, J = 8.0 Hz, 1 H), 7.75 (t, J = 7.5 Hz, 1 H), 7.39 (m, 13 H), 5.34 (s, 1 H), 5.29 (s, 1 H), 3.93 (s, 1 H); CIMS m/z (relative intensity) 474 (MH^+ , 100). Anal. Calcd for $\text{C}_{31}\text{H}_{23}\text{NO}_4$: C, H, N.

Example 26: *cis*-3-(3',4'-Dibenzylloxyphenyl)-4-carboxy-3,4-dihydro-*N*-methyl-6,7-dimethoxy-1-(2*H*)-isoquinolone (12h). 3,4-

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Dimethoxyhomophthalic anhydride (**10b**) (0.56 g, 2.5 mmol) was added to a stirred solution of 3,4-dibenzylxy benzylidenemethylamine (**11g**) (0.83 g, 2.5 mmol) in chloroform (3 mL). After 30 min, the yellow mixture became heterogeneous and ether was added to further precipitate the product. The light yellow precipitate was
5 collected and washed with chloroform to give a solid (0.59 g, 44%): mp 194-196°C; ¹H NMR (CDCl₃) δ 7.49 (s, 1 H), 7.34 (m, 11 H), 7.18 (s, 1 H), 6.91 (d, 1 H, J = 8.3 Hz), 6.79 (s, 1 H), 6.57 (d, 1 H, J = 8.3 Hz), 5.02 (s, 2 H), 4.98 (d, 1 H, J = 6.1 Hz), 4.92 (s, 2 H), 4.50 (d, 1 H, J = 5.8 Hz), 3.78 (s, 3 H), 3.74 (s, 3 H), 2.81 (s, 3 H); FABMS (*m*-NBA) *m/z* (relative intensity) 554 (MH⁺, 100).

10 **Example 27: 8,9-Dibenzylxy-5,6-dihydro-5,11-diketo-6-methyl-2,3-dimethoxy-11*H*-indeno[1,2-*c*] isoquinoline (13h)**. Thionyl chloride (15 mL) was added with stirring to the *cis* acid **12h** (1.2 g, 2.2 mmol). The result was an orange mixture that became dark red within 15 min. After 6 h, the reaction mixture was diluted with benzene (25 mL) and evaporated to dryness. Chloroform (7 mL)
15 was added to the purple solid and the solid was collected and washed with ether to give a light purple solid (0.75 g, 64%): mp 238-240°C; IR (thin film) 3027, 2963, 1685, 1649, 1493, 1458, 1252, 1203, 1088, 1014 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.96 (s, 1 H), 7.62 (s, 1 H), 7.38 (m, 1 H), 7.21 (s, 1 H), 7.11 (s, 1 H), 5.23 (d, 4 H, J = 5.2 Hz), 4.02 (s, 3 H) 3.95 (s, 3 H), 3.81 (s, 3 H); CIMS *m/z* (relative intensity) 534 (MH⁺, 22). Anal. Calcd for C₃₃H₂₇NO₆: C, H, N.

20 **Example 28: 3,4,5-Trimethoxybenzylidenemethylamine (11i)**. 3,4,5-Trimethoxybenzaldehyde (7.81 - 40.0 mmol) and a 40% aqueous solution of methylamine (20 mL) were stirred at room temperature for 2.5 h. The mixture was extracted with ether (4 x 75 mL), the ether layers were combined, and the solution
25 washed with saturated aqueous sodium chloride (75 mL), dried (MgSO₄), and concentrated under reduced pressure to give a colorless oil (7.94 g, 95%): IR (neat) 2940, 2840, 1646, 1576, 1500, 1453, 1407, 1369, 1323, 1230, 1115, 1013 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.18 (d, 1 H, J = 1.3 Hz), 6.95 (s, 2 H), 3.89 (s, 6 H), 3.87 (s, 3 H), 3.50 (d, 3 H, J = 1.3 Hz); CIMS *m/z* (relative intensity) 210 (MH⁺, 100).
30 Anal. Calcd for C₁₁H₁₅NO₃: C, H, N.

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Example 29: *cis*-4-Carboxy-3,4-dihydro-N-methyl-6,7-dimethoxy-3-(3',4',5'-trimethoxyphenyl)-1 (2*H*)isoquinolone (**12i**). 3,4-Dimethoxyhomophthalic anhydride (**10b**) (0.22 g, mmol) was added to a stirred solution of 3,4,5-trimethoxybenzylidenemethylamine (**11i**) (0.23 g, 1 mmol) in 5 chloroform (5 mL). After 30 min, the bright yellow homogeneous solution was tan and no solid was observed. Ether was added dropwise and the resulting precipitate was filtered and washed with ether to give fine white solid (0.1 g, 20%): mp, 229-231 °C; IR (neat) 2928, 1743, 1593, 1418, 1329, 1241, 1167, 1119 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 7.50 (s, 1 H), 7.15 (s, 1 H), 6.38 (s, 2 H), 5.0 (d, 1 H, *J* = 5.9 Hz), 4.48 (d, 1 H, *J* = 5.9 Hz), 3.79 (s, 3 H), 3.72 (s, 3 H), 3.59 (s, 9 H); CIMS *m/z* (relative intensity) 432 (MH⁺, 100). Anal. Calcd for C₂₂H₂₅NO₈: C, H, N.

Example 30: 5,6-Dihydro-5,11-diketo-6-methyl-2,3,8,9,10-pentamethoxy-11*H*-indeno[1,2]isoquinoline (**13i**). Thionyl chloride (15 mL) was added with stirring to the *cis* acid **12i** (1.2 g 2.8 mmol). The result was a yellow 15 mixture that became dark red within 15 min. After 4 h, the reaction mixture was diluted with benzene (25 mL) and evaporated to dryness. The purple solid was dissolved in chloroform and ether was added to give a precipitate that was collected and washed with ether to give a purple solid (0.75 g, 7.1 %): IR (neat) 2944, 1653, 1471, 1255, 1116, 1019 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.15 (s, 1 H), 7.69 (s, 1 H), 7.02 (s, 1 H), 4.11 (s, 3 H), 4.05 (s, 3 H), 4.02 (s, 3 H) 3.99 (s, 6 H), 3.91 (s, 3 H); 20 CIMS *m/z* (relative intensity) 412 (MH⁺, 100).

Example 31: *cis*-4-Carboxy-3,4-dihydro-N-methyl-3-(3',4',5'-trimethoxyphenyl)-1(2*H*)isoquinolone (**12j**). Homophthalic anhydride (**10a**) (0.32 g, 2 mmol) was added to a stirred solution of 3,4,5-trimethoxybenzylidenemethylamine (**11i**) (0.46 g, 2 mmol) in chloroform (5 mL). After 45 min, ether was added dropwise to the homogenous mixture and the resulting precipitate was filtered from the yellow solution and washed with ether to give a pale yellow solid (0.43 g, 60%): mp 194-195 °C; IR (neat) 2830, 1620, 1549, 1459, 1185, 1123 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.13 (s, 1 H), 7.99 (d, 1 H, *J* = 7.2 Hz), 7.52 (m, 4 H), 6.32 (s, 2 H), 5.04 (d, 1 H, *J* = 5.9 Hz), 4.63 (d, 1 H, *J* = 6.0 Hz), 3.58 (s, 3 H).

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H), 3.55 (s, 6 H), 2.94 (s, 3 H); CIMS *m/z* (relative intensity) 372 (MH⁺, 100). Anal. Calcd for C₂₀H₂₁NO₆: C, H, N.

Example 32: 5,6-Dihydro-5,11-diketo-6-methyl-8,9,10-trimethoxy-11*H*-indeno[1,2-c]isoquinoline (13j). Thionyl chloride (10 mL) was added with stirring to 12j (200 mg, 0.5 mmol). After 4 h, the reaction mixture was diluted with benzene (50 mL) and evaporated to dryness. The dark orange solid was dissolved in chloroform and ether was added to give a dark orange solid (16 mg, 10%): mp 194-195 °C; IR (neat) 2938, 1665, 1463, 1400, 1292, 1125, 1007, 976 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.67 (d, 1 H, *J* = 7.8 Hz), 8.32 (d, 1 H, *J* = 8.0 Hz), 7.68 (t, 1 H, *J* = 8.0 Hz), 7.45 (t, 1 H, *J* = 7.8 Hz), 7.04 (s, 1 H), 4.09 (s, 3 H), 4.06 (s, 3 H), 3.97 (s, 3 H), 3.89 (s, 3 H); CIMS *m/z* (relative intensity) 352 (MH⁺, 100). Anal. Calcd for C₂₀H₁₇NO₅: C, H, N.

Example 33: 3,4-Methylenedioxybenzylideneethylamine (11k). Piperonal (20.1 g, 0.14 mol) and a 70% aqueous solution of ethylamine (20 mL) were stirred at room temperature for 3 h. The mixture was extracted with ether (4 x 50 mL). The ether layers were combined and washed with aqueous sodium chloride (50 mL), dried (MgSO₄) and concentrated under reduced pressure to give a white crystalline powder (24.56 g, 93%): mp 47-48 °C; IR (KBr) 2963, 2836, 1645, 1603, 1498, 1480, 1441, 1252, 1092, 1031, 959, 926 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz) δ 8.15 (s, 1 H), 7.32 (d, 1 H, *J* = 1.3 Hz), 7.09 (dd, 1 H, *J* = 1.4, 6.0 Hz), 6.80 (d, 1 H, *J* = 8.0), 5.98 (s, 2 H) 3.59 (qd, 2 H, *J* = 6.0, 1.2 Hz), 1.26 (t, 3 H, *J* = 7.3 Hz); CIMS *m/z* (relative intensity) 178 (MH⁺, 100). Anal. Calcd for C₁₀H₁₁NO₂: C, H, N.

Example 34: *cis*-4-Carboxy-N-ethyl-3-(3',4'-methylenedioxyphenyl)-6,7-dimethoxy3,4-dihydro-1(2H)isoquinolone (12k). 3,4-Methylenedioxybenzylideneethylamine (11k) (0.89 g, 5.0 mmol) was stirred in chloroform (5.0 mL) and 4,5-dimethoxyhomophthalic anhydride (10b) (1.11 g, 5.0 mmol) was added. After 30 min, the yellow precipitate was filtered and washed with chloroform to give a pale yellow solid (0.58 g, 29 %): mp 231-233 °C (dec); IR (KBr) 2937, 1732, 1615, 1594, 1573, 1254, 1223, 1174, 1089, 1034, 986, 898 cm⁻¹; ¹H NMR (DMSO, 300 MHz) δ 7.50 (s, 1 H), 7.15 (s, 1 H), 6.76 (d, 1 H, *J* = 7.8 Hz), 6.57 (d, 1

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H, $J = 8.1$ Hz), 6.48 (s, 1 H), 5.94 (s, 2 H), 5.03 (d, 1 H, $J = 6.2$ Hz), 4.51 (d, 1 H, $J = 6.2$ Hz), 3.79 (dq, 1 H, $J = 6.9$ Hz), 3.80 (s, 3 H), 3.73 (s, 3 H), 2.96 (dq, 1 H, $J = 6.9$ Hz), 1.01 (t, 3 H, $J = 6.9$ Hz); FABMS (*m*-NBA) m/z (relative intensity) 400 (MH $^+$, 100).

5 **Example 35: 6-Ethyl-5,6-dihydro-5,11-diketo-2,3-dimethoxy-8,9-methylenedioxy-11*H*-indeno[1,2-*c*]isoquinoline (13k).** Thionyl chloride (6.0 mL) was added with stirring to the *cis* acid **12k** (0.58 g, 1.5 mmol) and the reaction mixture became dark reddish-purple and heterogeneous. After 4 h, the reaction mixture was diluted with benzene (5.0 mL) and evaporated to dryness. The brownish-red solid was loaded onto silica gel, passed through a short column of silica gel, eluting with chloroform, to give a brownish-red solid (0.34 g, 60%): mp 291-293 °C; IR 2969, 1694, 1643, 1613, 1555, 1486, 1393, 1308, 1252 cm $^{-1}$; 1 H NMR (CDCl $_3$, 300 MHz) δ 8.02 (s, 1 H), 7.65 (s, 1 H), 7.08 (s, 1 H), 7.01 (s, 1 H), 6.08 (s, 2 H), 4.49 (q, 2 H, $J = 7.2$ Hz), 4.03 (s, 3 H), 3.97 (s, 3 H), 1.50 (t, 3 H, $J = 7.2$ Hz); CIMS m/z (relative intensity) 366 (MH $^+$, 4.0). Anal. Calcd for C₂₁H₁₇NO₆: C, H, N.

10 15

Example 36: General Procedure for the Synthesis of Imines 17.

The *O*-TBDMS protected aminols **15** were synthesized using a reported procedure. The imines **17** were synthesized by treating the *O*-TBDMS protected aminols (9 mmol) with piperonal (9 mmol) in chloroform (20 mL) in the presence of anhydrous magnesium sulfate (2 g) at room temperature for 3 h. The imines were used as such for the next reaction without further purification. The crude yield of the imines **17** were quantitative.

20

Example 37: General Procedure for the Synthesis of Isoquinolones

18. 4,5-Dimethoxyhomophthalic anhydride (**10b**) (2.22 g, 10 mmol) was added to a 25 chloroform (20 mL) solution of the imine **17a** or **17b** (10 mmol) and the mixture was stirred at room temperature for 12 h, after which the TLC showed the complete disappearance of the starting materials and a white precipitate formed in the reaction mixture. The precipitated product was filtered off and washed with chloroform (5 mL) and dried to give pure **18a** or **18b**.

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Example 38: *cis-N-(t-Butyldimethylsilyloxybut-1-yl)-4-carboxy-3,4-dihydro-6,7dimethoxy-3-(3',4'-methylenedioxophenyl)-1(2H)isoquinolone (18a).* The isoquinolone **18a** was isolated in 36% yield: mp 239-240°C; IR (KBr) 3065, 2944, 1737 cm⁻¹; ¹H NMR (DMSO-d₆) δ 7.51 (s, 1 H), 7.11 (s, 1 H), 6.75 (d, *J* = 8.0 Hz, 1 H), 6.54 (dd, *J* = 1.3 and 8.1 Hz, 1 H), 6.46 (d, *J* = 1.2 Hz, 1 H), 5.93 (s, 2 H), 4.98 (d, *J* = 6.1 Hz, 1 H), 4.55 (d, *J* = 6.1 Hz, 1 H), 3.81 (s, 3 H), 3.80-8.70 (m, 1 H), 3.74 (s, 3 H), 3.53 (t, *J* = 5.78 Hz, 2 H), 2.95 - 2.80 (m, 1 H), 1.60 - 1.35 (m, 4 H), 0.88 (s, 9 H), - 0.98 (s, 6 H); ¹³C NMR (DMSO-d₆) δ 170.64, 162.47, 151.21, 147.65, 147.00, 146.81, 131.26, 126.84, 121.55, 121.43, 110.85, 109.81, 107.87, 107.71, 101.06, 62.20, 61.01, 55.44, 47.77, 45.25, 29.67, 29.54, 25.81, 24.07, 17.91, -5.37; CIMS *m/z* (relative intensity) 558 (MH⁺, 80). Anal. Calcd for C₂₉H₃₉NO₈Si: C, H, N.

Example 39: *cis-N-(t-Butyldimethylsilyloxypent-1-yl)-4-carboxy-3,4-dihydro-6,7dimethoxy-3-(3',4'-methylenedioxophenyl)-1(2H)isoquinolone (18b).* The isoquinolone **18b** was isolated in 57% yield: mp 240-242°C; IR (KBr) 3054, 2933, 1737 cm⁻¹; ¹H NMR (DMSO-d₆) δ 12.90 (bs, 1 H), 7.51 (s, 1 H), 7.09 (s, 1 H), 6.75 (d, *J* = 8.1 Hz, 1 H), 6.55 (dd, *J* = 1.6 and 8.1 Hz, 1 H), 6.47 (d, *J* = 1.3 Hz, 1 H), 5.93 (s, 2 H), 4.97 (d, *J* = 6.2 Hz, 1 H), 4.53 (d, *J* = 6.2 Hz, 1 H), 3.81 (s, 3 H), 3.83 - 8.70 (m, 1 H), 3.74 (s, 3 H), 3.52 (t, *J* = 6.2 Hz, 2 H), 2.85 - 2.73 (m, 1 H), 1.60 - 1.30 (m, 6 H), 0.82 (s, 9 H), - 0.99 (s, 6 H). CIMS *m/z* (relative intensity) 572 (MH⁺, 100). Anal. Calcd for C₃₀H₄₁NO₈Si: C, H, N.

Example 40: General Procedure for the Synthesis of Indenoisoquinolines 19. Thionyl chloride (10 mL) was added dropwise to the acid **18** (2 mmol) with stirring. The resulting solution was stirred at room temperature for 5 h after which the solution turned purple. Benzene (20 mL) was added to the reaction mixture and it was concentrated under reduced pressure. The resulting residue was passed through a short column of silica gel (230 - 400 mesh) eluting with chloroform:methanol (95:5). Concentration of the eluent resulted in a pink solid which was crystallized from ethyl acetate to obtain pure indenoisoquinolines **19**. Under the reaction conditions the deprotection of the *O*-TBDMS group was observed and only the hydroxy compounds were isolated.

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Example 41 :5,6-Dihydro-5,11-diketo-6-(4-hydroxybut-1-yl)-2,3-dimethoxy-8,9-methylenedioxy-(11H)indeno[1,2-c]isoquinoline (19a). The indenoisoquinoline 19a was isolated in 84% yield: mp 304-308 °C; IR (KBr) 3432, 2929, 1696, 1645, 1610 cm⁻¹; ¹H NMR (DMSO-d₆, 65 °C) δ 7.91 (s, 1 H), 7.53 (s, 1 H), 7.53 (s, 1 H), 7.06 (s, 1 H), 6.17 (s, 1 H), 4.43 (t, J = 7.7 Hz, 2 H), 3.90 (s, 3 H), 3.86 (s, 3 H), 3.45 (t, J = 5.8 Hz, 2 H), 1.88 - 1.70 (m, 2 H), 1.60-1.50 (m, 2 H); CIMS m/z (relative intensity) 424 (MH⁺, 100). Anal. Calcd for C₂₃H₂₁NO₇ 0.5 H₂O: C, H, N.

Example 42: 5,6-Dihydro-6-(4-hydroxypent-1-yl)-5,11-diketo-2,3-dimethoxy-8,9-methylenedioxy-11H-indenoisoquinoline (19b). The indenoisoquinoline 18b was isolated in 79% yield: mp 288-290 °C; IR (KBr) 3411, 2929, 1698, 1653, 1582, 1550 cm⁻¹; ¹H NMR (DMSO-d₆, 80 °C) δ 7.91 (s, 1 H), 7.53 (s, 1 H), 7.21 (s, 1 H), 7.07 (s, 1 H), 6.18 (s, 1 H), 4.41 (bs, 2 H), 3.90 (s, 3 H), 3.86 (s, 3 H), 3.60 (bs, 1 H), 3.40 (bs, 2 H), 1.88 - 1.70 (m, 2 H), 1.60 - 1.40 (m, 4 H); CIMS m/z (relative intensity) 438 (MH⁺, 100). Anal. Calcd for C₂₃H₂₁NO₇ 0.3 H₂O: C, H, N.

Example 43: cis-5,6,12,13-Tetrahydro-2,3-dimethoxy-6-methyl-5,11-dioxo-8,9(methylenedioxy)-(11H)indeno[1,2-c]isoquinoline (20). This compound was prepared as described previously in *J. Med. Chem.* 1984, 27, 544-547.

Example 44: cis-6-Ethyl-5,6,12,13-tetrahydro-2,3-dimethoxy-5,11-dioxo-8,9-(methylene-dioxy)-11H-indeno[1,2-c]isoquinoline (21). The acid 12k (3.99 g, 3 mmol) was added slowly under nitrogen to a solution of degassed Eaton's reagent (10% P₂O₅ in methanesulfonic acid, 120 mL) with stirring over a period of 20 min. The reaction mixture was stirred at room temperature for 4 h, after which the mixture was added dropwise to water (600 mL) with stirring. The precipitated white solid was filtered off and dissolved in chloroform (150 mL). The chloroform layer was washed with saturated NaHCO₃ solution (2 x 50 mL), water (50 mL), brine (60 mL) and dried (Na₂SO₄). Concentration of the organic layer gave the crude product, which was purified by column chromatography (4:1, hexane:ethyl acetate) to obtain pure 21 as a white solid (2.39 g, 63%). Neutralization of the bicarbonate layer with concd HCl gave the unreacted acid (0.821 g) as a white solid. Thus the yield based on

the recovered starting acid is 79.3%. An analytical sample was prepared by recrystallization from EtOAc-Hexane (1:1) to yield white prisms: mp 169-170°C; IR (KBr) 3006, 2994, 1706, 1642, 1601 cm⁻¹; ¹H NMR (CDCl₃) δ 7.59 (s, 1 H), 7.16 (s, 1 H), 7.06 (s, 1 H), 7.00 (s, 1 H), 6.09 (s, 1 H), 6.04 (s, 1 H), 5.04 (d, *J* = 6.9 Hz, 1 H), 5 4.70-4.53 (m, 1 H), 4.21 (d, *J* = 7.0 Hz, 1 H), 3.94 (s, 3 H), 3.88 (s, 3 H), 3.40-3.26 (m, 1 H), 1.35 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (CDCl₃) δ 198-8, 162.0, 154.7, 152.0, 150.6, 149.4, 148.5, 128.8, 126.4, 120.3, 110.2, 108.6, 104.2, 102.6, 56.6, 56.0, 55.8, 50.4, 43.3, 13.2. Anal. Calcd for C₂₁H₁₉NO₆: C, H, N.

Example 45: *cis*-6-Allyl-5,6,12,13-tetrahydro-2,3-dimethoxy-5,11-10 dioxo-8,9-(methylenedioxy)-(11*H*)indeno[1,2-*c*]isoquinoline (22).

Indenoisoquinoline **22** was synthesized in 72% yield from the acid **12c** in a similar procedure for the synthesis of indenoisoquinoline **21**. The treatment of the isoquinolone **12c** (4.11 g, 10 mmol) with Eaton's reagent (120 mL) provided the indenoisoquinoline **22** in 72% (2.83 g) yield: mp 178-180°C; IR (KBr) 2990, 1708, 15 1642, 1600 cm⁻¹; ¹H NMR (CDCl₃) δ 7.60 (s, 1 H), 7.17 (s, 1 H), 7.07 (s, 1 H), 7.03 (s, 1 H), 6.09 (s, 1 H), 6.05 (s, 1 H), 6.05 - 5.90 (m, 1 H), 5.45 - 5.20 (m, 3 H), 5.16 (d, *J* = 6.9 Hz, 1 H), 4.19 (d, *J* = 6.9 Hz, 1 H), 3.94 (s, 3 H), 3.88 (s, 3 H), 3.90 - 3.80 (m, 1 H); ¹³C NMR (CDCl₃) δ 198.8, 162.3, 154.8, 152.3, 150.6, 149.5, 148.6, 132.6, 129.0, 126.6, 120.0, 118.0, 110.4, 108.7, 104.4, 102.7, 102.6, 56.3, 56.1, 55.9, 50.3. 20 Anal. Calcd for C₂₂H₁₉NO₆: C, H, N.

Example 46: 5,6-Dihydro-5,11-diketo-2,3,8-trimethoxy-6-methyl-9[(methylsulfonyl)oxy]-(11*H*)indeno[1,2-*c*]isoquinoline (23). This compound was prepared as described previously in *J. Med. Chem.* 1985, 28, 1031-1036.

Example 47: 6-Ethyl-5,6,12α,13α-tetrahydro-11β-hydroxy-2,3-dimethoxy-8,9-(methylenedioxy)-5-oxo-11*H*-indeno[1,2-*c*]isoquinoline (24). The indenoisoquinoline **21** (0.381 g, 1 mmol) was heated at reflux with a 1 M solution of borane-tetrahydrofuran complex (4 mL) in dry THF (30 mL) for 1 h. After cooling, the reaction mixture was concentrated and the residue was dissolved in EtOAc (60 mL) and glacial acetic acid was added dropwise until pH 5. The organic layer was 30 washed with saturated sodium bicarbonate (2 x 50 mL), brine, and dried (Na₂SO₄) and

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concentrated. The residue on chromatographic purification (2% methanol in chloroform as eluent) provided the pure product **24** (0.363 g, 95%). An analytical sample was prepared by recrystallization from EtOAc-hexane (3:1) to yield white prisms: mp 189-192 °C; IR (KBr) 3468, 2919, 1630, 1594 cm⁻¹; ¹H NMR (CDCl₃) δ 7.62 (s, 1 H), 6.95 (s, 1 H), 6.94 (s, 1 H), 6.74 (s, 1 H), 5.99 (s, 1 H), 5.98 (s, 1 H), 4.97 (dd, *J* = 5.8 and 7.6 Hz, 1 H), 4.89 (d, *J* = 6.4 Hz, 1 H), 3.94 (s, 3 H), 3.93 (s, 3 H), 3.90-3.73 (m, 1 H), 3.59 (t, *J* = 5.8 Hz, 1 H), 3.45-3.30 (m, 1 H), 2.03 (d, *J* = 7.6 Hz, 1 H, D₂O exchangeable), 1.03 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (CDCl₃) δ 162.8, 152.1, 148.5, 148.4, 148.3, 138.7, 135.2, 128.2, 122.8, 110.5, 109.2, 106.6, 106.0, 101.5, 77.4, 60.0, 55.9, 55.8, 48.3, 37.9, 12.0. Anal. Calcd for C₂₁H₂₁NO₆: C, H, N.

Example 48: **6-Ethyl-5,6,12α,13α-tetrahydro-11β-hydroxy-2,3-dimethoxy-8,9-(methylenedioxy)-11*H*-indeno[1,2-c]isoquinoline (25).** The indenoisoquinoline **21** (2.391 g, 6.27 mmol) was heated at reflux with a 1M solution of borane-tetrahydrofuran complex (15 mL) in dry THF (100 mL) for 12 h. After cooling, the reaction mixture was concentrated and the residue was dissolved in EtOAc (100 mL) and glacial acetic acid was added dropwise until pH 5. The organic layer was washed with saturated sodium bicarbonate (2 X 100 mL), brine, and dried (Na₂SO₄) and concentrated. The residue on chromatographic purification (5% ethyl acetate in chloroform as eluent) provided the pure product **25** (2.13 g, 92%). An analytical sample was prepared by recrystallization from isopropanol to yield white crystals: mp 180-184 °C; IR (KBr) 3479, 2909, 1605, 1594 cm⁻¹; ¹H NMR (CDCl₃) δ 7.76 (s, 1 H), 6.90 (s, 1 H), 6.77 (s, 1 H), 6.66 (s, 1 H), 6.040 (s, 1 H), 6.02 (s, 1 H), 5.34 (dd, *J* = 3.1 and 6.6 Hz, 1 H), 4.90 (d, *J* = 8.4 Hz, 1 H), 4.15 (d, *J* = 16.2 Hz, 1 H), 4.06 (d, *J* = 16.2 Hz, 1 H), 3.93 (s, 3 H), 3.89 (s, 3 H) 371 (t, *J* = 7.5 Hz, 1 H), 2.86-2.70 (m, 1 H), 2.20-2.13 (m, 1 H), 2.03 (d, *J* = 3.1 Hz, 1 H, D₂O exchangeable), 1.02 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (CDCl₃) δ 149.4, 149.3, 148.4, 137.9, 131.2, 124.3, 123.4, 110.4, 110.1, 109.9, 104.7, 101.6, 74.0, 73.6, 58.0, 56.2, 56.0, 46.8, 45.6, 9.00. Anal. Calcd for C₂₁H₁₉NO₆.1.5 H₂O: C, H, N.

Example 49: **6-(3-Carboxy-1-propyl)-5,6-dihydro-5,11-diketo-11*H*-indeno[1,2-c]isoquinoline (26).** The indenoisoquinoline **7** (0.319 g, 1 mmol) was

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dissolved in acetone (50 mL) and cooled in an ice bath. Jones reagent was added dropwise to the cold solution of the alcohol until the red color of the reagent persisted. The excess Jones reagent was quenched by adding few drops of isopropyl alcohol. The reaction mixture was filtered through a small pad of celite and the residue was 5 washed with acetone (50 mL). The combined filtrate was concentrated and the residue was dissolved in saturated bicarbonate (100 mL) and the aqueous layer was washed with chloroform (2 x 30 mL). The aqueous layer was neutralized with concd HCl and extracted in CHCl₃ (3 x 50 mL). The combined organic layer was dried (Na₂SO₄) and concentrated to afford the acid as an orange solid. The solid was 10 crystallized from isopropyl alcohol to yield orange crystals (0.320 g, 96%): mp 204-206°C; IR (KBr) 3000 (b), 1708, 1698, 1654 cm⁻¹; ¹H NMR (CDCl₃) δ 8.68 (d, J = 8 Hz, 1 H), 8.30 (d, J = 8 Hz, 1 H), 7.86 (d, J = 7.4Hz, 1 H), 7.73 (t, J = 8.0 Hz, 1 H), 7.61 (d, J = 7.2 Hz, 1 H), 7.50 - 7.30 (m, 3 H), 4.60 (t, J = 7.8 Hz, 2 H), 3.71 (s, 1 H), 2.60 (t, J = 7.0 Hz, 2 H), 2.19 (p, J = 7.0 Hz, 2 H). Anal. Calcd for C₂₀H₁₅NO₄: C, H, 15 N.

Example 50: 6-Ethyl-2,3-dimethoxy-8,9-(methylenedioxy)-11H-indeno[1,2-c]isoquinolinium Chloride (27). The amino alcohol **25** (0.738 g, 2 mmol) was heated at reflux with 5% palladium on charcoal (0.265 g) in glacial acetic acid (100 mL) for 20 h. After cooling, the mixture was filtered through a small pad of 20 celite, and the solvent was evaporated to give a brown residue. The residue was dissolved in water (50 mL) and ethanol (6 mL) to give a light brown solution, to which was added 15% aqueous sodium chloride (10 mL). A yellow product precipitated immediately and was filtered, washed with ice cold water (10 mL), and dried over P₂O₅ under vacuum overnight to yield a yellow powder (0.552 g, 72%). An 25 analytical sample was crystallized from methanol: mp 340-343°C (dec); IR (KBr) 3382, 1480, 1305 and 1210 cm⁻¹; ¹H NMR (MeOH-d₄) δ 9.27 (s, 1 H), 7.61 (s, 2 H), 7.46 (s, 1 H), 7.30 (s, 1 H), 6.15 (s, 2 H), 5.03 (q, J = 7.2 Hz, 2 H), 4.87 (s, 2 H), 4.15 (s, 3 H), 4.05 (s, 3 H), 1.75 (t, J = 7.2 Hz, 3 H). ¹³C NMR (MeOH-d₄) δ 189.5, 162.4, 155.7, 155.0, 152.5, 147.3, 133.4, 130.8, 127.9, 123.6, 107.5, 107.3, 106.6, 105.4, 30 101.5, 57.7, 57.1, 54.8, 15.7. Anal. Calcd for C₂₁H₂₀NO₄Cl.H₂O: C, H, N.

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Example 51: Topoisomerase I-Mediated DNA Cleavage Reactions

Using 3'-End-labeled 161 BP Plasmid DNA. The 161 bp fragment from pBluescript SK(-) phagemid DNA (Stratagene, La Jolla, CA) was cleaved with the restriction endonuclease Pvu II and Hind III (New England Biolabs, Beverly, MA) in supplied NE buffer 2 (10 µL reactions) for 1 h at 37°C, separated by electrophoresis in a 1% agarose gel made in 1X TBE buffer. The 161 bp fragment was eluted from the gel slice (centrilutor by Amicon) and concentrated in a centricon 50 centrifugal concentrator (Amicon, Beverly, MA). Approximately 200 ng of the fragment was 3'-end-labeled at the Hind III site by fill-in reaction with [alpha-³²P]-dCTP and 0.5 mM dATP, dGTP, and dTTP, in React 2 buffer (50 mM Tris-HCl, pH 8.0, 100 mM MgCl₂, 50 mM NaCl) with 0.5 units of DNA polymerase I (Klenow fragment). Labeling reactions were followed by phenol-chloroform extraction and ethanol precipitation. The resulting 161 bp 3'-endlabeled DNA fragment was resuspended in water. Aliquots (approximately 50,000 dpm/reaction) were incubated with topoisomerase I at 30°C for 15 min in the presence of the indicated drug. Reactions were terminated by adding 0.5% SDS. After ethanol precipitation, the samples were resuspended in loading buffer (80% formamide, 10 mM sodium hydroxide, 1 mM sodium EDTA, 0.1% xylene cyanol, and 0.1% bromophenol blue, pH 8.0), and separated in a denaturing gel (16% polyacrylamide, 7 M urea) run at 51°C. The gel was dried and visualized by using a Phosphoimager and ImageQuant software (Molecular Dynamics, Sunnyvale, CA).

Example 52: Topoisomerase II-Mediated DNA Cleavage Assays

Using 5'-End-labeled Human C-myc DNA. A 403-base pair DNA fragment of the human c-myc gene from the junction between the first intron and the first exon was prepared by PCR between positions 2671 and 3073 using the oligonucleotides 5'-TGCCGCATCCACGAAACTTGC-3' as sense primer and 5'-GAACTGTTCAGTGTTCACCCCG-3' as antisense primer. Single-end labeling of these DNA fragments was obtained by 5'-end labeling of the adequate primer oligonucleotide. Approximately 0.1 µg of the human c-myc DNA that had been restricted by XhoI and XbaI was used as template for PCR. The 5'-end-labeled DNA

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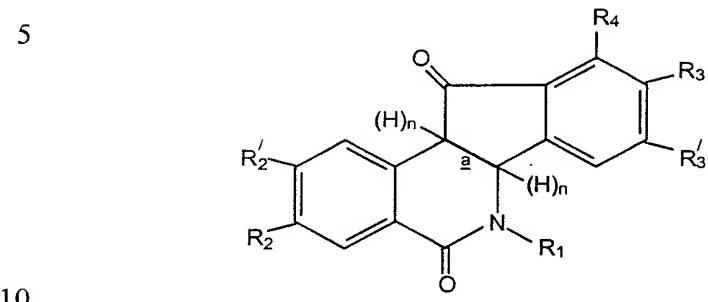
fragments were equilibrated with or without a drug in 1% dimethyl sulfoxide, 10 mM Tris-HCl, pH 7.5, 50 mM KC1, 5 mM MgCl₂, 2 mM dithiothreitol, 0.1 mM Na₂EDTA, 1 mM ATP, and 15 µg/mL bovine serum albumine for 5 min before addition of purified human topoisomerase II (40-70 ng) in a 10 µL final reaction volume. The reactions were performed at 37°C for 30 min and thereafter stopped by adding 1% sodium dodecyl sulfate (SDS) and 0.4 mg/mL proteinase K (final concentrations) followed by an additional incubation at 50°C for 30 min. Samples were ethanol-precipitated before separation of the topoisomerase II-cleaved fragments on denaturing polyacrylamide gels. The sequencing gels were made of 7% polyacrylamide in IX TBE buffer (90 mM Tris borate, 2 mM EDTA, pH 8.3). Electrophoresis was performed at 2500 V (60 W) for 2-5 h. The gels were dried and visualized using a Phosphoimager and ImageQuant software.

Example 53: SV40 DNA Unwinding Assay. Reaction mixtures (10 µL final volume) contained 0.3 µg supercoiled SV40 DNA in reaction buffer (10 mM Tris-HC1, pH 7.5, 50 mM KCl, 5 mM MgCl₂, 0.1 mM EDTA, 15 µg/mL bovine serum albumin) and 10 units of purified calf thymus topoisomerase I. Reactions were performed at 37°C for 30 min and terminated by the addition of 0.5% SDS, and then 1.1 µL of 10X loading buffer (20% Ficol 400, 0.1 M Na₂EDTA pH 8, 1.0% SDS. 0.25% Bromophenol Blue) was then added and reaction mixtures were loaded onto a 1% agarose gel made in IX TBE buffer. After electrophoresis, DNA bands were stained in 10 µg/mL of ethidium bromide and visualized by transillumination with UV light (300 nm).

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We claim:

1. A compound of formula:



wherein R₁ is hydrogen, formyl, phenyl, phenyl substituted with C₁-C₆ alkoxy or C₁-C₆ alkyl, or R₁ is a group -(CH₂)_mZ, wherein m is 1-6 and Z is selected from the group consisting of hydrogen, hydroxy, carboxy, formyl, C₁-C₆ alkyl, carbo-(C₁-C₆ alkoxy), C₂-C₆ alkenyl, phenyl, C₁-C₆ alkylamino, and C₁-C₆

15 hydroxyalkylamino;

R₂, R₂' and R₄ are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₂-C₆ alkenyl, and C₁-C₆ alkoxy, phenoxy and benzyloxy, or R₂ and R₂' taken together form a group of the formula -OCH₂O-;

20 R₃ and R₃' are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy, or R₃ and R₃' taken together form a group of the formula -OCH₂O-; and

25 wherein n = 1 or 0, and bond a is a single bond when n = 1, and bond a is a double bond when n = 0;

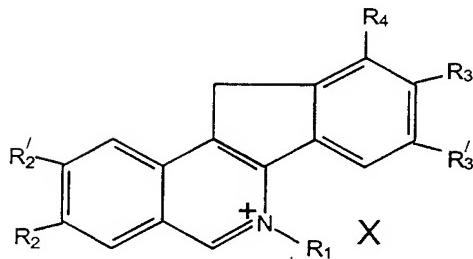
provided that when R₂, R₂', R₄, R₃ and R₃' are hydrogen, Z is not C₁-C₆ hydroxy alkylamino; and

further provided that when R₁ is methyl, R₃ and R₃' are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy.

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2. A compound of the formula:

5



10 wherein R₁ is phenyl or phenyl substituted with C₁-C₆ alkoxy or C₁-C₆ alkyl, or R₁ is a group -(CH₂)_mZ wherein m is 1-6 and Z is selected from the group consisting of hydrogen, hydroxy, carboxy, formyl, C₁-C₆ alkyl, carbo-(C₁-C₆ alkoxy), C₂-C₆ alkenyl, phenyl, C₁-C₆ alkylamino, and C₁-C₆ hydroxyalkylamino, provided that when Z is hydrogen, m is 2-6;

15 R₂, R₂' and R₄ are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₂-C₆ alkenyl, and C₁-C₆ alkoxy;

R₃ and R₃' are independently selected from the group consisting of hydrogen, C₁-C₆ alkyl, C₁-C₆ alkoxy, C₂-C₆ alkenyl, phenoxy, and benzyloxy, or R₃ and R₃' taken together form a group of the formula -OCH₂O-; and

20 wherein X is a pharmaceutically acceptable anion.

3. A pharmaceutical composition for treatment of cancer comprising a compound of claim 1 in an amount effective for treatment of said cancer, and a pharmaceutically acceptable carrier, excipient or diluent therefore.

4. A method for treating a patient having cancer, said method comprising the step of administering to the patient an effective amount of a compound according to claim 1.

5. A pharmaceutical composition for treatment of cancer comprising a compound of claim 2 in an amount effective for treatment of said cancer, and a pharmaceutically acceptable carrier, excipient or diluent therefore.

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6. A method for treating a patient having cancer, said method comprising the step of administering to the patient an effective amount of a compound according to claim 2.

7. The compound of claim 1 wherein R₁ is ethyl, allyl, -(CH₂)₃COOH or 5 -(CH₂)_mOH wherein m=4or 5;

R₂ and R₂' are independently selected from hydrogen and methoxy;

R₃ and R₃' are hydrogen or together form the group -OCH₂O-; and

R₄ is hydrogen.

8. A pharmaceutical composition for treatment of cancer comprising a 10 compound of claim 7 in an amount effective for treatment of said cancer, and a pharmaceutically acceptable carrier, excipient or diluent therefore.

9. A method for treating a patient having cancer, said method comprising the step of administering to the patient an effective amount of a compound according to claim 7.

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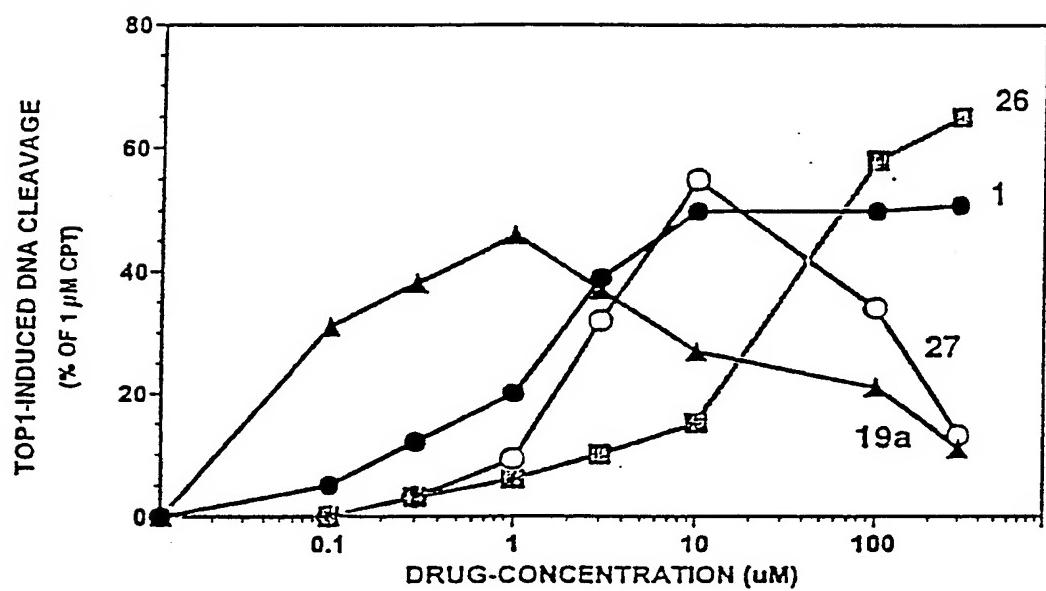


Figure 1.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/23900

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) :Please See Extra Sheet.
US CL :514/280, 284; 546/48, 61
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/280, 284; 546/48, 61

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

CAS ONLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 5,597,831 A (MICHALSKY, deceased, et al.) 28 January 1997, see entire document, especially column 1 and columns 5-6, Examples 1-2.	1, 3, 4
X	PEREZ-CHIESA et al. Evaluation of genotoxicity of the indenoisoquinoline analogues of fagaronine and nitidine in Drosophila melanogaster. Mutation Research. 1993, Vol. 301, pages 207-212, especially page 208.	2, 5, 6
X	WAWZONEK et al. The synthesis and reactions of 1-carbamyl-11-ketoindeno[1,2-c]isoquinoline. J. Org. Chem. April 1966, Vol. 31, pages 1004-1006, especially page 1004, compounds IIa, b.	1

Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"B" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

14 DECEMBER 1999

Date of mailing of the international search report

03 FEB 2000

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/23900

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	JAYARAMAN et al. Novel oxidative transformation of indenoisoquinolines to isoquinoline-3-spiro-3'-phthalides in the presence of osmium tetroxide and 4-methylmorpholine N-oxide. J. Org. Chem. 21 August 1998, Vol. 63, No. 17, pages 572-573, especially page 572, compounds 7b, e.	1, 7
X	Chem. abstr., Vol 96, No. 23, 07 June 1982 (Columbus. OH, USA), pages 653-654, column 2, the abstract No. 199485x, WAWZONEK, S. 'Synthesis of 6-substituted-6-H-indeno[1, 2-c]isoquinoline-5,11-diones.' Org. Prep. Proced. Int. 1982, 14(3), 163-8.	1, 7

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/23900

A. CLASSIFICATION OF SUBJECT MATTER:
IPC (7):

A61K 31/473, 31/4741; C07D 221/18